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חוק הפטנטים, תשכ"ז-1967
PATENT LAW, 5727-1967

בקשה לפטנט
Application for Patent

מספר: Number	114615
תאריך: Date	16-07-1995
הוקדם/נרחה Ante/Post-dated	

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רחובות

Assignment בעל אמצאה מכח העברה
of an invention the title of which is Owner, by virtue of

מודולטורים של התפקוד של קולטני FAS וחלבונים אחרים (בעברית)
(Hebrew)

MODULATORS OF THE FUNCTION OF FAS RECEPTORS AND OTHER PROTEINS (באנגלית)
(English)

hereby apply for a patent to be granted to me in respect thereof.

מבקש בואת כי ינתן לי עליה פטנט

• בקשת חלוקה • Application of Division		• בקשת פטנט מוסף • Application for Patent Addition		• דרישה ריב קדימה • Priority Claim		
מבקשת פטנט from Application	לבקשה/לפטנט to Patent/Appl.	מספר/סימן Number/Mark	תאריך Date	מדינת האגוד Convention Country		
No. dated	No. dated					
• יפוי כח: אסמ'א/אסמ'א / עוד יוגש P.O.A.: to be filed later- filed in case הוגש בענין						
חשון למסירת מסמכים בישראל מספר: Y/94-84C Address for Service in Israel הנרי עינב, עורך פטנטים איגטר-לאר בע"מ, קריית ויצמן נס-ציונה 76110						
חתימת המבקש Signature of Applicant For the Applicant Henry Einav		היום 16 בחודש יולי שנת 1995 This 16 of July 1995 of the year				



לשימוש הלשכה
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מודולטורים של התפקוד של קולטני FAS וחלבונים אחרים

**MODULATORS OF THE FUNCTION OF FAS
RECEPTORS AND OTHER PROTEINS**

Field of the Invention

The present invention is generally in the field of receptors belonging to the TNF/NGF superfamily of receptors and the control of their biological functions. The TNF/NGF superfamily of receptors includes receptors such as the p55 and p75 tumor necrosis factor receptors (TNF-Rs) and the FAS ligand receptor (also called FAS/APO1 or FAS-R and hereinafter will be called FAS-R) and others. More specifically, the present invention concerns a novel protein, herein designated MORT-1 (also called HF-1) which binds to the intracellular domain (IC) of the Fas-R, (this intracellular domain designated Fas-IC) and which novel protein is capable of modulating the function of the Fas-R. Further, MORT-1 is also capable of self-association and can activate cell cytotoxicity on its own. Accordingly, the present invention also concerns new proteins capable of binding to MORT-1 which are capable of modulating the function of MORT-1. The present invention also concerns the preparation and uses of MORT-1.

It should be noted that HF-1 (the original designation) and MORT-1 (the presently used designation) are both used throughout the specification and denote the same protein.

Background of the Invention and Prior Art

This application is related to application nos. 112002 and 112692 in which there are described, amongst other aspects, the isolation and characterization of the protein HF1 (or MORT-1), which binds specifically to the FAS-IC. In these earlier applications, there are also described other FAS-IC-binding proteins such as, for example, those designated F2, F9 and DD11, as well as the FAS-IC itself which is capable of self-association. The present application, as detailed herein below, concerns MORT-1 in general, which is distinguished from the other abovementioned FAS-IC-binding proteins in that it binds only to the FAS-IC whilst these other proteins also bind to the intracellular domain of the p55 TNF-R (p55-IC). Moreover, the present invention concerns specifically additional characteristics and uses of MORT-1 or analogs, fragments and derivatives thereof, as well as additional characteristics and uses of one of the new MORT-1-binding proteins.

Tumor Necrosis Factor (TNF- α) and Lymphotoxin (TNF- β) (hereinafter, TNF, refers to both TNF- α and TNF- β) are multifunctional pro-inflammatory cytokines formed mainly by mononuclear phagocytes, which have many effects on cells (Wallach, D. (1986) in: Interferon 7 (Ion Gresser, ed.), pp. 83-122, Academic Press, London; and Beutler and Cerami (1987)). Both TNF- α and TNF- β initiate their effects by binding to specific cell surface receptors. Some of the effects are likely to be beneficial to the organism : they may destroy, for example tumor cells or virus infected cells and augment antibacterial activities of granulocytes. In this way, TNF contributes to the defense of the organism against tumors and infectious agents and contributes to the recovery from injury. Thus, TNF can be used as an anti-tumor agent in which application it binds to its receptors on the surface of tumor cells and thereby initiates the events leading to the death of the tumor cells. TNF can also be used as an anti-infectious agent.

However, both TNF- α and TNF- β also have deleterious effects. There is evidence that over-production of TNF- α can play a major pathogenic role in several diseases. Thus, effects of TNF- α , primarily on the vasculature, are now known to be a major cause for symptoms of septic shock (Tracey et al., 1986). In some diseases, TNF may cause excessive loss of weight (cachexia) by suppressing activities of adipocytes and by causing anorexia, and TNF- α was thus called cachetin. It was also described as a mediator of the damage to tissues in rheumatic diseases (Beutler and Cerami, 1987) and as a major mediator of the damage observed in graft-versus-host reactions (Piquet et al., 1987). In addition, TNF is known to be involved in the process of inflammation and in many other diseases.

Two distinct, independently expressed, receptors, the p55 and p75 TNF-Rs, which bind both TNF- α and TNF- β specifically, initiate and/or mediate the above noted biological effects of TNF. These two receptors have structurally dissimilar intracellular domains suggesting that they signal differently (See Hohmann et al., 1989; Engelmann et al., 1990; Brockhaus et al., 1990; Leotscher et al., 1990; Schall et al., 1990; Nophar et al., 1990; Smith et al., 1990; and Heller et al., 1990). However, the cellular mechanisms, for example, the various proteins and possibly other factors, which are involved in the intracellular signaling of the p55 and p75 TNF-Rs have yet to be elucidated (In IL 109632 and as set forth also

herein below, there is described for the first time, new proteins capable of binding to the p75IC and p55 IC). It is this intracellular signaling, which occurs usually after the binding of the ligand, i.e. TNF (α or β), to the receptor, that is responsible for the commencement of the cascade of reactions that ultimately result in the observed response of the cell to TNF.

As regards the above mentioned cytotoxic effect of TNF, in most cells studied so far, this effect is triggered mainly by the p55 TNF-R. Antibodies against the extracellular domain (ligand binding domain) of the p55 TNF-R can themselves trigger the cytotoxic effect (see EP 412486) which correlates with the effectivity of receptor cross-linking by the antibodies, believed to be the first step in the generation of the intracellular signaling process. Further, mutational studies (Brakebusch et al., 1992; Tartaglia et al., 1993) have shown that the biological function of the p55 TNF-R depends on the integrity of its intracellular domain, and accordingly it has been suggested that the initiation of intracellular signaling leading to the cytotoxic effect of TNF occurs as a consequence of the association of two or more intracellular domains of the p55 TNF-R. Moreover, TNF (α and β) occurs as a homotrimer and as such has been suggested to induce intracellular signaling via the p55 TNF-R by way of its ability to bind to and to cross-link the receptor molecules, i.e. cause receptor aggregation. In IL 109632 and IL 111125 and also herein below there is described how the p55IC and p55DD can self-associate and induce, in a ligand-independent fashion, TNF-associated effects in cells.

Another member of the TNF/NGF superfamily of receptors is the FAS receptor (FAS-R) which has also been called the Fas antigen, a cell-surface protein expressed in various tissues and sharing homology with a number of cell-surface receptors including TNF-R and NGF-R. The FAS-R mediates cell death in the form of apoptosis (Itoh et al., 1991), and appears to serve as a negative selector of autoreactive T cells, i.e. during maturation of T cells, FAS-R mediates the apoptotic death of T cells recognizing self-antigens. It has also been found that mutations in the FAS-R gene (*lpr*) cause a lymphoproliferation disorder in mice that resembles the human autoimmune disease systemic lupus erythematosus (SLE) (Watanabe-Fukunaga et al., 1992). The ligand for the FAS-R appears to be a cell-surface associated molecule carried by, amongst others, killer T cells (or cytotoxic T lymphocytes -

CTLs), and hence when such CTLs contact cells carrying FAS-R, they are capable of inducing apoptotic cell death of the FAS-R-carrying cells. Further, a monoclonal antibody has been prepared that is specific for FAS-R, this monoclonal antibody being capable of inducing apoptotic cell death in cells carrying FAS-R, including mouse cells transformed by cDNA encoding human FAS-R (Itoh et al., 1991).

It has also been found that various other normal cells, besides T lymphocytes, express the FAS-R on their surface and can be killed by the triggering of this receptor. Uncontrolled induction of such a killing process is suspected to contribute to tissue damage in certain diseases, for example, the destruction of liver cells in acute hepatitis. Accordingly, finding ways to restrain the cytotoxic activity of FAS-R may have therapeutic potential.

Conversely, since it has also been found that certain malignant cells and HIV-infected cells carry the FAS-R on their surface, antibodies against FAS-R, or the FAS-R ligand, may be used to trigger the FAS-R mediated cytotoxic effects in these and thereby provide a means for combating such malignant cells or HIV-infected cells (see Itoh et al., 1991). Finding yet other ways for enhancing the cytotoxic activity of FAS-R may therefore also have therapeutic potential.

It has been a long felt need to provide a way for modulating the cellular response to TNF (α or β) and FAS-R ligand, for example, in pathological situations as mentioned above, where TNF or FAS-R ligand is over-expressed it is desirable to inhibit the TNF- or FAS-R ligand- induced cytotoxic effects, while in other situations, e.g. wound healing applications, it is desirable to enhance the TNF effect, or in the case of FAS-R, in tumor cells or HIV-infected cells it is desirable to enhance the FAS-R mediated effect.

A number of approaches have been made by the present inventors (see for example, European Application Nos. EP-186833, EP 308378, EP 398327 and EP 412486) to regulate the deleterious effects of TNF by inhibiting the binding of TNF to its receptors using anti-TNF antibodies or by using soluble TNF receptors (being essentially the soluble extracellular domains of the receptors) to compete with the binding of TNF to the cell surface-bound TNF-Rs. Further, on the basis that TNF-binding to its receptors is required for the TNF-induced cellular effects, approaches by the present inventors (see for example IL 101769 and

its corresponding EPO 568925) have been made to modulate the TNF effect by modulating the activity of the TNF-Rs. Briefly, EPO 568925 (IL 101769) relates to a method of modulating signal transduction and/or cleavage in TNF-Rs whereby peptides or other molecules may interact either with the receptor itself or with effector proteins interacting with the receptor, thus modulating the normal functioning of the TNF-Rs. In EPO 568925 there is described the construction and characterization of various mutant p55 TNF-Rs, having mutations in the extracellular, transmembranal, and intracellular domains of the p55 TNF-R. In this way regions within the above domains of the p55 TNF-R were identified as being essential to the functioning of the receptor, i.e. the binding of the ligand (TNF) and the subsequent signal transduction and intracellular signaling which ultimately results in the observed TNF-effect on the cells. Further, there is also described a number of approaches to isolate and identify proteins, peptides or other factors which are capable of binding to the various regions in the above domains of the TNF-R, which proteins, peptides and other factors may be involved in regulating or modulating the activity of the TNF-R. A number of approaches for isolating and cloning the DNA sequences encoding such proteins and peptides; for constructing expression vectors for the production of these proteins and peptides; and for the preparation of antibodies or fragments thereof which interact with the TNF-R or with the above proteins and peptides that bind various regions of the TNF-R, are also set forth in EPO 568925. However, EPO 568925 does not specify the actual proteins and peptides which bind to the intracellular domains of the TNF-Rs (e.g. p55 TNF-R), nor does it describe the yeast two-hybrid approach to isolate and identify such proteins or peptides which bind to the intracellular domains of TNF-Rs. Similarly, heretofore there has been no disclosure of proteins or peptides capable of binding the intracellular domain of FAS-R.

Thus, when it is desired to inhibit the effect of TNF, or the FAS-R ligand, it would be desirable to decrease the amount or the activity of TNF-Rs or FAS-R at the cell surface, while an increase in the amount or the activity of TNF-Rs or FAS-R would be desired when an enhanced TNF or FAS-R ligand effect is sought. To this end the promoters of both the p55 TNF-R and the p75 TNF-R have been sequenced, analyzed and a number of key

sequence motifs have been found that are specific to various transcription regulating factors, and as such the expression of these TNF-Rs can be controlled at their promoter level, i.e. inhibition of transcription from the promoters for a decrease in the number of receptors, and an enhancement of transcription from the promoters for an increase in the number of receptors (see IL 104355 and IL 109633). Corresponding studies concerning the control of FAS-R at the level of the promoter of the FAS-R gene have yet to be reported.

Further, it should also be mentioned that, while it is known that the tumor necrosis factor (TNF) receptors, and the structurally-related receptor FAS-R, trigger in cells, upon stimulation by leukocyte-produced ligands, destructive activities that lead to their own demise, the mechanisms of this triggering are still little understood. Mutational studies indicate that in FAS-R and the p55 TNF receptor (p55-R) signaling for cytotoxicity involve distinct regions within their intracellular domains (Brakebusch et al., 1992; Tartaglia et al., 1993; Itoh and Nagata, 1993). These regions (the 'death domains') have sequence similarity. The 'death domains' of both FAS-R and p55-R tend to self-associate. Their self-association apparently promotes that receptor aggregation which is necessary for initiation of signaling (see IL 109632, IL 111125 and IL 112002, as well as Song et al., 1994; Wallach et al., 1994; Boldin et al., 1995) and at high levels of receptor expression can result in triggering of ligand-independent signaling (IL 109632, IL 111125 and Boldin et al., 1995).

Thus, prior to IL 109632 and the present invention, there have not been provided proteins which may regulate the effect of ligands belonging to the TNF/NGF superfamily, such as the TNF or FAS-R ligand effect on cells, by mediation of the intracellular signaling process, which signaling is probably governed to a large extent by the intracellular domains (ICs) of the receptors belonging to the TNF/NGF superfamily of receptors, such as those of the TNF-Rs, i.e. the p55 and p75 TNF-R intracellular domains (p55IC and p75IC, respectively), as well as the FAS-IC.

Accordingly, it is one aim of the invention to provide proteins, being MORT-1, analogs, fragments or derivatives thereof, which are capable of binding to the intracellular domain of the FAS-R, which proteins are presently believed to be involved in the intracellular signaling process initiated by the binding of FAS ligand to its receptor. The MORT-1

protein, analogs, fragments and derivatives thereof of the present invention are distinct from the FAS-IC-binding proteins described in the earlier co-pending applications

Another aim of the invention is to provide antagonists (e.g. antibodies) to these FAS-IC binding molecules, being the MORT-1 protein, analogs fragments and derivatives, which may be used to inhibit the signaling process, when desired, when such FAS-IC-binding proteins are positive signal effectors (i.e. induce signaling), or to enhance the signaling process, when desired, when such FAS-IC-binding proteins are negative signal effectors (i.e. inhibit signaling).

Yet another aim of the invention is to use such MORT-1 protein, analogs, fragments and derivatives, to isolate and characterize additional proteins or factors, which may, for example, be involved further downstream in the signaling process, and/or to isolate and identify other receptors further upstream in the signaling process to which these MORT-1 protein, analogs, fragments and derivatives bind (e.g. other FAS-Rs or related receptors), and hence, in whose function they are also involved.

Moreover, it is an aim of the present invention to use the above-mentioned MORT-1 protein, analogs, fragments and derivatives as antigens for the preparation of polyclonal and/or monoclonal antibodies thereto. The antibodies, in turn, may be used, for example, for the purification of the new MORT-1 protein from different sources, such as cell extracts or transformed cell lines.

Furthermore, these antibodies may be used for diagnostic purposes, e.g. for identifying disorders related to abnormal functioning of cellular effects mediated by the FAS-R receptor.

A further aim of the invention is to provide pharmaceutical compositions comprising the above MORT-1 protein, analogs, fragments or derivatives, as well as pharmaceutical compositions comprising the above noted antibodies or other antagonists.

Summary of the Invention

In accordance with the present invention, we have found a novel protein which is capable of binding to the intracellular domain of the FAS-R. This FAS-IC-binding protein

may act as a mediator or modulator of the FAS-R ligand effect on cells by way of mediating or modulating the intracellular signaling process which usually occurs following the binding of the FAS-R ligand at the cell surface.

This novel protein has been designated HF1, or MORT-1 (for 'Mediator of Receptor Toxicity'), and in addition to its FAS-IC-binding specificity has other characteristics (see Example 1), for example, it has a region homologous to the 'death domain' (DD) regions of the p55-TNF-R and FAS-R (p55-DD and FAS-DD), and thereby is also capable of self-association. MORT-1 is also capable of activating cell cytotoxicity on its own, an activity possibly related to its self-association capability. It has now also been found that co-expression of the region in MORT-1 (HF1) that contains the 'death domain' homology sequence (MORT-DD, present in the C-terminal part of MORT-1) strongly interferes with FAS-induced cell death, as would be expected from its ability to bind to the 'death domain' of the FAS-IC. Further, in the same experimental conditions it was found that co-expression of the part of MORT-1 that does not contain the MORT-DD region (the N-terminal part of MORT-1, amino acids 1-117, 'MORT-1 head') resulted in no interference of the FAS-induced cell death and, if at all, a somewhat enhanced FAS-induced cell cytotoxicity.

Accordingly, it is likely that MORT-1 also binds to other proteins involved in the intracellular signaling process. These MORT-1-binding proteins may therefore also act as indirect mediators or modulators of the FAS-R ligand effect on cells by way of mediating or modulating the activity of MORT-1; or these MORT-1-binding proteins may act directly as mediators or modulators of the MORT-1-associated intracellular signaling process by way of mediating or modulating the activity of MORT-1, which, as noted above, has an apparently independent ability to activate cell cytotoxicity. These MORT-1-binding proteins may also be used in any of the standard screening procedures to isolate, identify and characterize additional proteins, peptides, factors, antibodies, etc. which may be involved in the MORT-1-associated or FAS-R-associated signaling process or may be elements of other intracellular signaling processes. One such MORT-1-binding protein has been isolated and is described herein.

Accordingly, the present invention provides a DNA sequence encoding a herein designated MORT-1 protein, analogs, or fragments thereof, all of which are capable of binding to or interacting with the intracellular domain of the FAS-ligand receptor (FAS-IC).

In particular, the present invention provides a DNA sequence selected from the group consisting of :

- (a) a cDNA sequence derived from the coding region of a native MORT-1 protein;
- (b) DNA sequences capable of hybridization to a cDNA of (a) under moderately stringent conditions and which encode a biologically active FAS-R intracellular domain-binding protein; and
- (c) DNA sequences which are degenerate as a result of the genetic code to the DNA sequences defined in (a) and (b) and which encode a biologically active FAS-R intracellular domain-binding protein.

A specific embodiment of the above DNA sequence of the invention is a DNA sequence encoding the protein MORT-1 comprising the sequence depicted in Fig. 4.

The present invention also provides a MORT-1 protein, analogs, fragments or derivatives thereof encoded by any of the above sequences of the invention, said proteins, analogs, fragments and derivatives being capable of binding to or interacting with the intracellular domain of the FAS-R.

A specific embodiment of the above protein of the invention is the MORT-1 protein having the deduced amino acid sequence depicted in Fig. 4.

Also provided by the present invention are vectors encoding the above MORT-1 protein, analogs, fragments or derivatives of the invention, which contain the above DNA sequence of the invention, these vectors being capable of being expressed in suitable eukaryotic or prokaryotic host cells; transformed eukaryotic or prokaryotic host cells containing such vectors; and a method for producing the MORT-1 protein, analogs, fragments or derivatives of the invention by growing such transformed host cells under conditions suitable for the expression of said protein, analogs, fragments or derivatives,

effecting post-translational modifications of said protein as necessary for obtention of said protein and extracting said expressed protein, analogs, fragments or derivatives from the culture medium of said transformed cells or from cell extracts of said transformed cells.

In another aspect, the present invention also provides antibodies or active derivatives or fragments thereof specific to the MORT-1 protein, analogs, fragments and derivatives thereof, of the invention.

By yet another aspect of the invention, there are provided various uses of the above DNA sequences or the proteins which they encode, according to the invention, which uses include amongst others :

- (i) a method for the modulation of the FAS-R ligand effect on cells carrying a FAS-R, comprising treating said cells with one or more MORT-1 protein, analogs, fragments or derivatives, according to the invention, all of which being capable of binding to the intracellular domain and modulating the activity of said FAS-R, wherein said treating of the cells comprises introducing into said cells said one or more MORT-1 protein, analogs, fragments or derivatives in a form suitable for intracellular administration or introducing into said cells, a DNA sequence encoding said one or more proteins, analogs, fragments or derivatives in the form of a suitable expression vector carrying said sequence, said vector being capable of effecting the insertion of said sequence into said cells in the way that said sequence is expressed in said cells;
- (ii) a method for modulating the FAS-R ligand effect on cells comprising treating said cells with MORT-1, analogs, fragments or derivatives thereof, all being capable of binding to the intracellular domain and modifying the activity of FAS-R wherein said treating of cells comprises introducing into said cells said MORT-1, analogs, fragments or derivatives in a form suitable for intracellular introduction thereof, or introducing into said cells a DNA sequence encoding said MORT-1, analogs, fragments or derivatives in the form of a suitable vector carrying said sequence, said vector being capable of effecting the insertion of said sequence into said cells in a way that said sequence is expressed in said cells.

- (iii) a method as in (ii) above wherein said treating of said cells is by transfection of said cells with a recombinant animal virus vector comprising the steps of :
- (a) constructing a recombinant animal virus vector carrying a sequence encoding a viral surface protein (ligand) that is capable of binding to a specific cell surface receptor on the surface of a FAS-R-carrying cell and a second sequence encoding a protein selected from the MORT-1 protein, analogs, fragments and derivatives of the invention, that when expressed in said cells is capable of modulating the activity of said FAS-R; and
 - (b) infecting said cells with said vector of (a).
- (iv) a method for modulating the FAS-R ligand effect on cells carrying a FAS-R comprising treating said cells with antibodies or active derivatives or fragments thereof according to the invention, said treating being by application of a suitable composition containing said antibodies, active fragments or derivatives thereof to said cells, wherein when the MORT-1 proteins or portions thereof of said cells are exposed on the extracellular surface, said composition is formulated for extracellular application, and when said MORT-1 proteins are intracellular said composition is formulated for intracellular application;
- (v) a method for modulating the FAS-R ligand effect on cells carrying a FAS-R comprising treating said cells with an oligonucleotide sequence encoding an antisense sequence of at least part of the MORT-1 sequence of the invention, said oligonucleotide sequence being capable of blocking the expression of the MORT-1 protein;
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- (vi) a method as in (v) above wherein said treating of cells is by transfection of said cells with a recombinant animal virus vector comprising the steps of :
- (a) constructing a recombinant animal virus vector carrying a sequence encoding a viral surface protein (ligand) that is capable of binding to a specific cell surface receptor on the surface of a FAS-R-carrying cell and a second sequence which is an oligonucleotide sequence encoding an antisense sequence of at least part of the MORT-1 sequence according to the invention, said oligonucleotide sequence being

capable of blocking the expression of the MORT-1 protein when introduced into said cells by said virus; and

(b) infecting said cells with said vector of (a).

(vii) a method for treating tumor cells or HIV-infected cells, or other diseased cells, comprising:

(a) constructing a recombinant animal virus vector carrying a sequence encoding a viral surface protein that is capable of binding to a tumor cell surface receptor or HIV-infected cell surface receptor or a receptor carried by other diseased cells and a sequence encoding a protein selected from the MORT-1 protein, analogs, fragments and derivatives of the invention, that when expressed in said tumor, HIV-infected cell, or other diseased cell is capable of killing said cell; and

(b) infecting said tumor or HIV-infected cells or other diseased cells with said vector of (a).

(viii) a method for modulating the FAS-R ligand effect on cells comprising applying the ribozyme procedure in which a vector encoding a ribozyme sequence capable of interacting with a cellular mRNA sequence encoding a MORT-1 protein of the invention is introduced into said cells in a form that permits expression of said ribozyme sequence in said cells, and wherein when said ribozyme sequence is expressed in said cells it interacts with said cellular mRNA sequence and cleaves said mRNA sequence resulting in the inhibition of expression of said MORT-1 protein in said cells;

~~(ix) a method selected from any of the above methods wherein said MORT-1 protein or said MORT-1 encoding sequence comprises at least that part of the MORT-1 protein which binds specifically to the FAS-IC, or at least that part of the MORT-1 encoding sequence that encodes that part of the MORT-1 protein which binds specifically to the FAS-IC;~~

(x) a method for isolating and identifying proteins, factors or receptors capable of binding to the MORT-1 protein according to the invention, comprising applying the procedure of affinity chromatography in which said MORT-1 protein is attached to

the affinity chromatography matrix, said attached protein is brought into contact with a cell extract; and proteins, factors or receptors from the cell extract which bound to said attached MORT-1 protein are then eluted, isolated and analyzed;

- (xi) a method for isolating and identifying proteins, capable of binding to the MORT-1 protein according to the invention, comprising applying the yeast two-hybrid procedure in which a sequence encoding said MORT-1 protein is carried by one hybrid vector and a sequence from a cDNA or genomic DNA library is carried by the second hybrid vector, the vectors then being used to transform yeast host cells and the positive transformed cells being isolated, followed by extraction of the said second hybrid vector to obtain a sequence encoding a protein which binds to said MORT-1 protein; and
- (xii) a method for isolating and identifying a protein capable of binding to the intracellular domain of FAS-R comprising applying the procedure of non-stringent southern hybridization followed by PCR cloning, in which a sequence or parts thereof according to the invention is used as a probe to bind sequences from a cDNA or genomic DNA library, having at least partial homology thereto, said bound sequences then amplified and cloned by the PCR procedure to yield clones encoding proteins having at least partial homology to said sequences according to the invention.

The present invention also provides a pharmaceutical composition for the modulation of the FAS ligand- effect on cells comprising, as active ingredient, any one of the following :

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- (i) a MORT-1 protein according to the invention, its biologically active fragments, analogs, derivatives or mixtures thereof; (ii) a recombinant animal virus vector encoding a protein capable of binding a cell surface receptor and encoding a MORT-1 protein or its biologically active fragments or analogs according to the invention; and (iii) an oligonucleotide sequence encoding an antisense sequence of the MORT-1 sequence of the invention, wherein said oligonucleotide sequence may be the second sequence of the recombinant animal virus vector of (ii) above.

It should be mentioned that MORT-1 has a distinct region which binds to the FAS-IC and another distinct region which is involved in self-association of MORT-1, and accordingly, these distinct regions or parts thereof may be used independently to identify other proteins, receptors, etc. which are capable of binding to MORT-1 or to FAS-R and which may be involved in the MORT-1- or FAS-R- related intracellular signaling processes. Further, MORT-1 may have other activities associated with either of the above distinct regions or other regions of MORT-1 or combinations thereof, for example, enzymatic activity, which may be related to the cell cytotoxic effects of MORT-1 on its own. Thus, MORT-1 may also be used to specifically identify other proteins, peptides, etc. which may be involved in such additional activities associated with MORT-1.

Another aspect of the invention concerns proteins which are capable of binding specifically to MORT-1. As set forth in Example 1 below, MORT-1 is capable of self-association and of activating cell cytotoxicity on its own. Thus, MORT-1 may also bind to other proteins (besides FAS-R) which are involved in the intracellular signaling process. These MORT-1 binding proteins may act either as modulators of the MORT-1-associated intracellular signaling process or as modulators of the FAS-R ligand effect by modulating the activity of MORT-1.

Accordingly, the present invention also provides a DNA sequence encoding a protein capable of binding to MORT-1. In particular, the above MORT-1-binding protein sequence is selected from the group consisting of a cDNA sequence derived from a coding region of a native MORT-1-binding protein sequence; DNA sequences capable of hybridizing to the above cDNA sequence under moderately stringent conditions and which encode a biologically active MORT-1-binding protein; and DNA sequences which are degenerate as a result of the genetic code to the above cDNA and DNA sequences and which encode a biologically active MORT-1-binding protein. A specific embodiment of the above DNA sequence of the invention is a DNA sequence encoding a MORT-1-binding protein comprising the partial sequence set forth in Fig. 8.

Further, there is also provided a MORT-1-binding protein, analogs, fragments, or derivatives thereof encoded by any of the above sequences, said MORT-1-binding proteins, analogs, fragments and derivatives being capable of binding to MORT-1.

Moreover, there are also provided: vectors encoding the above MORT-1-binding proteins, which contain the MORT-1-binding protein encoding sequences, these vectors being capable of being expressed in suitable eukaryotic or prokaryotic host cells; transformed eukaryotic or prokaryotic host cells containing such vectors; and a method for producing the MORT-1-binding proteins, analogs, fragments or derivatives thereof, by growing such transformed host cells under conditions suitable for the expression of the MORT-1-binding protein, analogs, fragments or derivatives thereof, effecting post-translational modifications of the MORT-1-binding protein as necessary for obtention of said protein and extracting said expressed MORT-1-binding protein, analogs, fragments or derivatives from the culture medium of said transformed cells or from cell extracts of said transformed cells; and antibodies or active derivatives or fragments thereof specific to the MORT-1-binding proteins, analogs or derivatives thereof.

Furthermore, the above DNA sequences encoding the MORT-1-binding proteins, analogs or derivatives thereof, or the proteins, analogs and derivatives which they encode, may be used in a number of ways (see uses (i)-(xii) above) for the specific modulation of the MORT-1-associated effect on cells or the FAS-R effect on cells by way of modulation of the MORT-1 binding to FAS-R.

The isolation of the MORT-1-binding proteins, their identification and characterization may be carried out by any of the standard screening techniques used for isolating and identifying proteins, for example, the yeast two-hybrid method, affinity chromatography methods, and any of the other well-known standard procedures used for this purpose.

Other aspects and embodiments of the present invention are also provided as arising from the following detailed description of the invention.

It should be noted that, where used throughout, the following terms : "Modulation of the FAS-ligand effect on cells"; and "Modulation of the MORT-1 effect on cells" are understood to encompass *in vitro* as well as *in vivo* treatment.

Brief Description of the Drawings

Fig. 1 A and B are reproductions of autoradiograms of SDS-PAGE gels (10% acrylamide) showing the interaction between HF1 (MORT1) and FAS-IC *in vitro*. Fig. 1A shows a control autoradiogram of an immunoprecipitate of the proteins (from extracts of HeLa cells transfected with the FLAG-HF1 (FLAG-MORT1) fusion protein or with the luciferase cDNA (control), the immunoprecipitation being performed with anti-FLAG antibody; and

Fig. 1B shows an autoradiogram of a representative gel performed to evaluate the *in vitro* interaction between HF1 and FAS-IC by way of assessing, autoradiographically, the binding of [³⁵S]-methionine-metabolically labeled HF1 produced in transfected HeLa cells as a fusion protein with the FLAG octapeptide (FLAG-MORT1) to GST, human and mouse GST-FAS-IC fusion protein (GST-huFAS-IC, GST-mFAS-IC) and GST-FAS-IC fusion proteins in which the FAS-IC contained an Ile to Ala replacement mutation at position 225 (GST-mFAS-IC I225A). The [³⁵S]labeled proteins of the HeLa cells including the labeled FLAG-MORT1 fusion protein having been first extracted were subjected to interaction with the various GST and GST-FAS-IC proteins (bound to glutathione beads) and then to SDS-PAGE. As controls in all of the interaction experiments, extracts of HeLa cells transfected with luciferase were subjected to interactions with the GST and GST-FAS-IC fusion proteins and SDS-PAGE. Figs. 1 A and B are also described in Example 1.

Fig. 2 A, B and C are reproductions of autoradiograms of SDS-PAGE gels (10% acrylamide) on which were separated various immunoprecipitates from transfected HeLa cells and which show the *in vivo* interaction of HF1 (MORT1) with FAS-IC. The HeLa cells were transfected with DNA constructs encoding : HF1-FLAG (FLAG-MORT1) fusion protein alone, HF1-FLAG fusion protein and the human

FAS-R (FLAG-MORT1 + Fas/APO1) or human FAS-R alone (Fas/APO1) (Fig. 2A); or with HF1-FLAG fusion protein and the human p55R (FLAG-MORT1 + p55R) (Fig. 2B); or with HF1-FLAG fusion protein and a chimeric fusion protein between human FAS-R and p55-R in which the extracellular domain of the FAS-R was replaced with the corresponding region of the p55-R (FLAG-MORT1 + p55-FAS chimera) or the FAS-R-p55-R chimeric fusion protein alone (p55-Fas chimera) (Fig. 2C). In all cases the transfected cells were metabolically labeled with [35 S]cysteine (20 μ Ci/ml) and [35 S]methionine (40 μ Ci/ml), and were subjected to protein extraction. The protein extracts from the different transfected cells were then immunoprecipitated with various antibodies being anti-FLAG, anti-FAS, anti-p75-R and anti-p55-R antibodies (α FLAG, α FAS, α p75-R and α p55-R in Figs. 2A-C) and subjected to SDS-PAGE. On the left side of Fig. 2A there is indicated the protein bands corresponding to FAS-R (Fas/APO1) and HF1-FLAG (FLAG-MORT1); between Figs. 2A and B are shown the relative positions of standard molecular weight markers (in kDa); and on the right hand side of Fig. 2C the protein bands corresponding to p55R and the p55-FAS chimera are indicated. Figs. 2A-C are also described in Example 1.

Fig. 3 shows a reproduction of a Northern blot in which poly A⁺ RNA (0.3 μ g) from HeLa cells transfected was probed with HF1 cDNA, as described in Example 1.

Figs. 4 depicts schematically the preliminary nucleotide and deduced amino acid sequence of HF1, as described in Example 1, in which the 'death domain' is underlined as is a possible translation start site, i.e. the underlined methionine residue at position 49 (bold, underlined M). The asterisk indicates the translation stop codon (nucleotides 769-771). At the beginning and in the middle of each line are provided two numerals depicting the relative positions of the nucleotides and amino acids of the sequence with respect to the start of the sequence (5' end), in which the first numeral denotes the nucleotide and the second numeral denoted the amino acid.

Fig. 5 shows the results of experiments to determine the C-terminal end of MORT-1, wherein Fig. 5 is a reproduction of an autoradiogram of an SDS-PAGE gel (10%

acrylamide) on which were separated various MORT-1-FLAG fusion products expressed in HeLa cells and metabolically labeled with ^{35}S -cysteine and ^{35}S -methionine followed by immunoprecipitation with either anti-FLAG monoclonal antibodies (M2) (lanes 2, 4 and 6) or as a control, anti-p75 TNF-R antibodies (#9) (lanes 1, 3 and 5), as described in Example 1.

Figs. 6 (A and B) depict graphically the ligand-independent triggering of cytotoxic effects in cells transfected with MORT-1, wherein cell viability was determined either by the neutral red uptake assay (Fig. 6A), or for specifically determining the viability of cells that expressed the transfected DNA, by measuring the amounts of placental alkaline phosphatase secreted into the medium (Fig. 6B). HeLa cells were transfected with tetracycline-controlled expression vectors encoding HF1 (MORT1), human FAS-IC, human p55-IC, or luciferase (control), and in all cases also with a cDNA encoding the secreted alkaline phosphatase, which permitted the evaluation of the effect of transient expression of these proteins on the viability of the cells. In both Figs. 6A and 6B the open graphs represent transfected cells grown in the presence of tetracycline (1 $\mu\text{g}/\text{ml}$, to block expression) and the closed graphs represent transfected cells grown in the absence of tetracycline. Figs. 6A and B are also described in Example 1.

Fig. 7 is a graphic representation of the effects of different portions of the MORT-1 protein on the cell cytotoxic effects mediated by FAS-R, as described in Example 1.

Fig. 8 is a schematic presentation of the preliminary partial nucleotide sequence encoding a MORT-1-binding protein, obtained from a cDNA clone as described in Example 2.

Detailed Description of the Invention

The present invention relates, in one aspect, to a novel protein MORT-1 (HF1) which is capable of binding to the intracellular domain of the FAS-R receptor, a member of the TNF/NGF superfamily and hence is considered as a mediator or modulator of FAS-R, having a role in, for example, the signaling process that is initiated by the binding of FAS ligand to FAS-R. The amino acid and DNA sequences of MORT-1 according to the invention also

represent new sequences; they do not appear in the 'GENEBANK' or 'PROTEIN BANK' data banks of DNA or amino acid sequences.

Thus, the present invention concerns the DNA sequence encoding the MORT-1 protein and the MORT-1 protein encoded by the DNA sequences.

Moreover, the present invention also concerns the DNA sequences encoding biologically active analogs, fragments and derivatives of the MORT-1 protein, and the analogs, fragments and derivatives encoded thereby. The preparation of such analogs, fragments and derivatives is by standard procedure (see for example, Sambrook et al., 1989) in which in the DNA sequences encoding the MORT-1 protein, one or more codons may be deleted, added or substituted by another, to yield analogs having at least a one amino acid residue change with respect to the native protein. Acceptable analogs are those which retain at least the capability of binding to the intracellular domain of the FAS-R, or which can mediate any other binding or enzymatic activity, e.g. analogs which bind the FAS-IC but which do not signal, i.e. do not bind to a further downstream receptor, protein or other factor, or do not catalyze a signal-dependent reaction. In such a way analogs can be produced which have a so-called dominant-negative effect, namely, an analog which is defective either in binding to the FAS-IC, or in subsequent signaling following such binding. Such analogs can be used, for example, to inhibit the FAS-ligand- effect by competing with the natural FAS-IC-binding proteins. Likewise, so-called dominant-positive analogs may be produced which would serve to enhance the FAS ligand effect. These would have the same or better FAS-IC-binding properties and the same or better signaling properties of the natural FAS-IC-binding proteins. In an analogous fashion, biologically active fragments of MORT-1

may be prepared as noted above with respect to the analogs of MORT-1. Suitable fragments of MORT-1 are those which retain the FAS-IC binding capability or which can mediate any other binding or enzymatic activity as noted above. Accordingly, MORT-1 fragments can be prepared which have a dominant-negative or a dominant-positive effect as noted above with respect to the analogs. It should be noted that these fragments represent a special class of the analogs of the invention, namely, they are defined portions of MORT-1 derived from the full MORT-1 sequence, each such portion or fragment having any of the above-noted desired

activities. Similarly, derivatives may be prepared by standard modifications of the side groups of one or more amino acid residues of the MORT-1 protein, its analogs or fragments, or by conjugation of the MORT-1 protein, its analogs or fragments, to another molecule e.g. an antibody, enzyme, receptor, etc., as are well known in the art.

The new MORT-1 protein, its analogs, fragments and derivatives have a number of possible uses, for example:

- (i) They may be used to mimic or enhance the function of FAS-R ligand, in situations where an enhanced FAS-R ligand effect is desired such as in anti-tumor, anti-inflammatory or anti-HIV applications where the FAS-R ligand- induced cytotoxicity is desired. In this case the MORT-1 protein, its analogs, fragments or derivatives, which enhance the FAS-R ligand effect, i.e. cytotoxic effect, may be introduced to the cells by standard procedures known per se. For example, as the MORT-1 protein is intracellular and it should be introduced only into the cells where the FAS-R ligand effect is desired, a system for specific introduction of this protein into the cells is necessary. One way of doing this is by creating a recombinant animal virus e.g. one derived from Vaccinia, to the DNA of which the following two genes will be introduced: the gene encoding a ligand that binds to cell surface proteins specifically expressed by the cells e.g. ones such as the AIDs (HIV) virus gp120 protein which binds specifically to some cells (CD4 lymphocytes and related leukemias) or any other ligand that binds specifically to cells carrying a FAS-R, such that the recombinant virus vector will be capable of binding such FAS-R- carrying cells; and the gene encoding the MORT-1 protein. Thus, expression of the cell-surface-binding

protein on the surface of the virus will target the virus specifically to the tumor cell or other FAS-R- carrying cell, following which the MORT-1 protein encoding sequence will be introduced into the cells via the virus, and once expressed in the cells will result in enhancement of the FAS-R ligand effect leading to the death of the tumor cells or other FAS-R- carrying cells it is desired to kill. Construction of such recombinant animal virus is by standard procedures (see for example, Sambrook et al., 1989). Another possibility is to introduce the sequences of the MORT-1 protein

in the form of oligonucleotides which can be absorbed by the cells and expressed therein.

- (ii) They may be used to inhibit the FAS-R ligand effect, e.g. in cases such as tissue damage in septic shock, graft-vs.-host rejection, or acute hepatitis, in which it is desired to block the FAS-R ligand induced FAS-R intracellular signaling. In this situation it is possible, for example, to introduce into the cells, by standard procedures, oligonucleotides having the anti-sense coding sequence for the MORT-1 protein, which would effectively block the translation of mRNAs encoding the MORT-1 protein and thereby block its expression and lead to the inhibition of the FAS-R ligand- effect. Such oligonucleotides may be introduced into the cells using the above recombinant virus approach, the second sequence carried by the virus being the oligonucleotide sequence.

Another possibility is to use antibodies specific for the MORT-1 protein to inhibit its intracellular signaling activity.

Yet another way of inhibiting the FAS-R ligand effect is by the recently developed ribozyme approach. Ribozymes are catalytic RNA molecules that specifically cleave RNAs. Ribozymes may be engineered to cleave target RNAs of choice, e.g. the mRNAs encoding the MORT-1 protein of the invention. Such ribozymes would have a sequence specific for the MORT-1 mRNA and would be capable of interacting therewith (complementary binding) followed by cleavage of the mRNA, resulting in a decrease (or complete loss) in the expression of the MORT-1 protein, the level of decreased expression being dependent upon the level of ribozyme expression in the target cell. To introduce ribozymes into the cells of choice (e.g. those carrying FAS-R) any suitable vector may be used, e.g. plasmid, animal virus (retrovirus) vectors, that are usually used for this purpose (see also (i) above, where the virus has, as second sequence, a cDNA encoding the ribozyme sequence of choice). (For reviews, methods etc. concerning ribozymes see Chen et al., 1992; Zhao and Pick, 1993; Shore et al., 1993; Joseph and Burke, 1993; Shimayama et al.,

1993; Cantor et al., 1993; Barinaga, 1993; Crisell et al., 1993 and Koizumi et al., 1993).

- (iii) They may be used to isolate, identify and clone other proteins which are capable of binding to them, e.g. other proteins involved in the intracellular signaling process that are downstream of the TNF-R or FAS-R intracellular domain. For example, the MORT-1 protein, namely, the DNA sequence encoding it may be used in the yeast two-hybrid system (see Example 1, below) in which the sequence of the MORT-1 protein will be used as "bait" to isolate, clone and identify from cDNA or genomic DNA libraries other sequences ("preys") encoding proteins which can bind to the MORT-1 protein. In the same way, it may also be determined whether the MORT-1 protein of the present invention can bind to other cellular proteins, e.g. other receptors of the TNF/NGF superfamily of receptors.
- (iv) The MORT-1 protein, its analogs, fragments or derivatives may also be used to isolate, identify and clone other proteins of the same class i.e. those binding to FAS-R intracellular domain or to functionally related receptors, and involved in the intracellular signaling process. In this application the above noted yeast two-hybrid system may be used, or there may be used a recently developed system employing non-stringent southern hybridization followed by PCR cloning (Wilks et al., 1989). In the Wilks et al. publication, there is described the identification and cloning of two putative protein-tyrosine kinases by application of non-stringent southern hybridization followed by cloning by PCR based on the known sequence of the kinase motif, a conceived kinase sequence. This approach may be used, in accordance with the present invention using the sequence of the MORT-1 protein to identify and clone those of related FAS-R intracellular domain-binding proteins.
- (v) Yet another approach to utilizing the MORT-1 protein, its analogs, fragments or derivatives of the invention is to use them in methods of affinity chromatography to isolate and identify other proteins or factors to which they are capable of binding, e.g. other receptors related to FAS-R or other proteins or factors involved in the intracellular signaling process. In this application, the MORT-1 protein, its analogs,

fragments or derivatives of the present invention, may be individually attached to affinity chromatography matrices and then brought into contact with cell extracts or isolated proteins or factors suspected of being involved in the intracellular signaling process. Following the affinity chromatography procedure, the other proteins or factors which bind to the MORT-1 protein, its analogs, fragments or derivatives of the invention, can be eluted, isolated and characterized.

- (vi) As noted above, the MORT-1 protein, its analogs, fragments or derivatives of the invention may also be used as immunogens (antigens) to produce specific antibodies thereto. These antibodies may also be used for the purposes of purification of the MORT-1 protein either from cell extracts or from transformed cell lines producing MORT-1, its analogs or fragments. Further, these antibodies may be used for diagnostic purposes for identifying disorders related to abnormal functioning of the FAS-R ligand system, e.g. overactive or underactive FAS-R ligand- induced cellular effects. Thus, should such disorders be related to a malfunctioning intracellular signaling system involving the MORT-1 protein, such antibodies would serve as an important diagnostic tool.
- (vii) MORT-1 may also be used as an indirect modulator of a number of other proteins by virtue of its capability of binding to other intracellular proteins, (the so-called MORT-1 binding proteins, see below), which other intracellular proteins directly bind yet other intracellular proteins or an intracellular domain of a transmembrane protein. An example of such a protein or such an intracellular domain is the well-known p55 TNF receptor, the intracellular signalling of which is modulated by a number of proteins which bind directly to its intracellular domain (see copending IL 109632). In fact we have isolated such a MORT-1 binding protein (see below and Example 2) which binds to the intracellular domain of the p55 TNF receptor.

For the purposes of modulating these other intracellular proteins or the intracellular domains of transmembranal proteins, MORT-1 may be introduced into cells in a number of ways as mentioned hereinabove in (ii).

It should also be noted that the isolation, identification and characterization of the MORT-1 protein of the invention may be performed using any of the well known standard screening procedures. For example, one of these screening procedures, the yeast two-hybrid procedure as is set forth herein (Example 1), was used to identify the MORT-1 protein of the invention. Likewise as noted above and below, other procedures may be employed such as affinity chromatography, DNA hybridization procedures, etc. as are well known in the art, to isolate, identify and characterize the MORT-1 protein of the invention or to isolate, identify and characterize additional proteins, factors, receptors, etc. which are capable of binding to the MORT-1 protein of the invention.

Furthermore, it should also be noted that amongst the characteristics of MORT-1 is its ability to bind to the FAS-IC and also its ability to self-associate. MORT-1 is also capable of activating cell cytotoxicity on its own, an activity related to its self-association ability. It appears (see Example 1) that the part of MORT-1 which binds to the FAS-IC is distinct from the part of MORT-1 which is involved in its self-association. MORT-1 may also have other activities which may be a function of the above noted distinct parts of the MORT-1 molecule or other parts of the molecule or combinations of any of these parts. These other activities may be enzymatic or related to binding other proteins (e.g. MORT-1-binding proteins or other receptors, factors, etc.). Thus, MORT-1 may be used in the above methods for modulation of FAS-R-ligand effects or its own MORT-1-mediated cellular effects, or it may be used in the modulation of other cellular signaling processes related to other receptors, factors, etc.

~~More specifically, by this aspect of the invention the MORT-1 encoding DNA~~
molecule itself and mutations thereof (ie. encoding analogs or active fractions of MORT-1) can be used for gene therapy (i.e. by the ways set forth in uses (i) and (ii) above) for modulating the activity of the FAS-R (or modulating or mediating the FAS ligand-effect on cells). Moreover, as MORT-1 also has a cytotoxic effect on cells, these MORT-1 or mutant MORT-1 encoding DNA molecules may also be used for gene therapy for modulating the MORT-1 effect in cells (also by way of the uses (i) and (ii) above). In these gene therapy applications, the MORT-1, analogs or derivatives may be used in three ways:

(a) the whole MORT-1 protein, its analogs, derivatives or active fragments which have both FAS-IC and MORT-1-binding ability (i.e. contain the two regions of MORT-1, one of which is involved in binding to FAS-IC and the other which is involved in the self-association of MORT-1) may be used to modulate FAS-R and MORT-1-associated effects;

(b) the part of MORT-1, and analogs, derivatives, and active fragments of this part which binds to the FAS-IC may be used for inducing a 'dominant negative' effect on FAS-IC, i.e. inhibition of FAS-R-mediated cellular effects, or may be used for inducing a 'gain of function' effect on FAS-IC, i.e. enhancement of the FAS-R-mediated cellular effects; and

(c) the part of MORT-1 and analogs, derivatives and active fractions of this part, which binds specifically to MORT-1 may be used for induction of 'dominant negative' or 'gain of function' effects on MORT-1, i.e. either inhibition or enhancement of MORT-1-associated cellular effects.

As set forth in uses (iii)-(v) above, the DNA sequence encoding MORT-1 or the MORT-1 protein may be used to isolate, identify and characterize other proteins which are capable of specifically binding to MORT-1, i.e. the MORT-1-binding proteins of the invention. For example, the MORT-1 encoding DNA molecule may be used with the yeast two-hybrid technique to isolate, identify and characterize MORT-1-binding proteins; or the MORT-1 protein may be used by affinity chromatography procedures to isolate such MORT-1-binding proteins. Furthermore, any of the other well known standard methods of the art for isolating, identifying and characterizing proteins may be employed for isolating, identifying and characterizing the MORT-1-binding proteins, in which methods the MORT-1, or analogs, derivatives or fragments thereof or DNA sequences encoding any of the aforementioned may be used, where appropriate, to isolate, identify and subsequently to characterize the MORT-1-binding proteins.

The MORT-1-binding proteins may be produced as mentioned hereinabove by any of the standard recombinant DNA procedures, as may be produced analogs, derivatives or active fragments of these MORT-1-binding proteins.

The MORT-1-binding proteins may be utilized, along the lines set forth in the uses (i) and (ii) above to mimic or enhance the MORT-1-associated cellular effects or to inhibit the

MORT-1-associated cellular effects. Further, the MORT-1-binding proteins may also be used, where applicable, to enhance or to inhibit, indirectly, the FAS-R ligand-induced effects by way of enhancing or inhibiting the activity of MORT-1.

Furthermore, these MORT-1 binding proteins may also bind to the intracellular domain of other receptor proteins belonging to the TNF/NGF superfamily or to other intracellular proteins, and thereby be involved in the modulation of these intracellular domains or intracellular proteins. One such MORT-1 binding protein has been isolated and it was found that it also binds to the intracellular domain of the p55 TNF-R. Hence this MORT-1 binding protein appears to be involved in the modulation of p55 TNF-R.

As set forth in use (vi) above, the MORT-1 protein or MORT-1-binding proteins may be used to generate antibodies specific to MORT-1 or MORT-1-binding proteins. These antibodies or fragments thereof may be used as set forth hereinbelow in detail, it being understood that in these applications the antibodies or fragments thereof are those specific for MORT-1 or MORT-1-binding proteins.

Moreover, the MORT-1-binding proteins may themselves be employed, in an analogous fashion to the way in which MORT-1 was used as noted above and below, to isolate, identify and characterize other proteins, factors, etc. which are capable of binding to the MORT-1-binding proteins and which may represent factors involved further downstream in the associated signaling process, or which may be able to bind MORT-1 or FAS-R or other receptors or factors and thereby also being involved in the associated signaling process, or which may have cell cytotoxic effects on their own (like MORT-1) and hence would represent proteins involved in a distinct signaling process. In this regard, it should also be mentioned that the MORT-1-binding proteins may also have enzymatic or cell cytotoxic functions and thus may also be involved in distinct signaling processes, permitting the use of the MORT-1-binding proteins and/or MORT-1 and/or further proteins, factors, receptors, etc. which bind to any of MORT-1 or MORT-1-binding proteins in methods for modulating these MORT-1-binding protein-associated enzymatic or cell cytotoxic functions.

As regards the antibodies mentioned herein throughout, the term "antibody" is meant to include polyclonal antibodies, monoclonal antibodies (mAbs), chimeric antibodies, anti-

idiotypic (anti-Id) antibodies to antibodies that can be labeled in soluble or bound form, as well as fragments thereof provided by any known technique, such as, but not limited to enzymatic cleavage, peptide synthesis or recombinant techniques.

Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of animals immunized with an antigen. A monoclonal antibody contains a substantially homogeneous population of antibodies specific to antigens, which populations contains substantially similar epitope binding sites. MAbs may be obtained by methods known to those skilled in the art. See, for example Kohler and Milstein, *Nature*, 256:495-497 (1975); U.S. Patent No. 4,376,110; Ausubel et al., eds., Harlow and Lane ANTIBODIES : A LABORATORY MANUAL, Cold Spring Harbor Laboratory (1988); and Colligan et al., eds., Current Protocols in Immunology, Greene publishing Assoc. and Wiley Interscience N.Y., (1992, 1993), the contents of which references are incorporated entirely herein by reference. Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, GILD and any subclass thereof. A hybridoma producing a mAb of the present invention may be cultivated *in vitro*, *in situ* or *in vivo*. Production of high titers of mAbs *in vivo* or *in situ* makes this the presently preferred method of production.

Chimeric antibodies are molecules different portions of which are derived from different animal species, such as those having the variable region derived from a murine mAb and a human immunoglobulin constant region. Chimeric antibodies are primarily used to reduce immunogenicity in application and to increase yields in production, for example, where murine mAbs have higher yields from hybridomas but higher immunogenicity in humans, such that human/murine chimeric mAbs are used. Chimeric antibodies and methods

for their production are known in the art (Cabilly et al., *Proc. Natl. Acad. Sci. USA* 81:3273-3277 (1984); Morrison et al., *Proc. Natl. Acad. Sci. USA* 81:6851-6855 (1984); Boulianne et al., *Nature* 312:643-646 (1984); Cabilly et al., European Patent Application 125023 (published November 14, 1984); Neuberger et al., *Nature* 314:268-270 (1985); Taniguchi et al., European Patent Application 171496 (published February 19, 1985); Morrison et al., European Patent Application 173494 (published March 5, 1986); Neuberger et al., PCT Application WO 8601533, (published March 13, 1986); Kudo et al., European Patent

Application 184187 (published June 11, 1986); Sahagan et al., *J. Immunol.* 137:1066-1074 (1986); Robinson et al., International Patent Application No. WO8702671 (published May 7, 1987); Liu et al., *Proc. Natl. Acad. Sci USA* 84:3439-3443 (1987); Sun et al., *Proc. Natl. Acad. Sci USA* 84:214-218 (1987); Better et al., *Science* 240:1041-1043 (1988); and Harlow and Lane, *ANTIBODIES :A LABORATORY MANUAL*, supra. These references are entirely incorporated herein by reference.

An anti-idiotypic (anti-Id) antibody is an antibody which recognizes unique determinants generally associated with the antigen-binding site of an antibody. An Id antibody can be prepared by immunizing an animal of the same species and genetic type (e.g. mouse strain) as the source of the mAb with the mAb to which an anti-Id is being prepared. The immunized animal will recognize and respond to the idiotypic determinants of the immunizing antibody by producing an antibody to these idiotypic determinants (the anti-Id antibody). See, for example, U.S. Patent No. 4,699,880, which is herein entirely incorporated by reference.

The anti-Id antibody may also be used as an "immunogen" to induce an immune response in yet another animal, producing a so-called anti-anti-Id antibody. The anti-anti-Id may be epitopically identical to the original mAb which induced the anti-Id. Thus, by using antibodies to the idiotypic determinants of a mAb, it is possible to identify other clones expressing antibodies of identical specificity.

Accordingly, mAbs generated against the MORT-1 protein, or MORT-1-binding proteins, analogs, fragments or derivatives thereof, of the present invention may be used to induce anti-Id antibodies in suitable animals, such as BALB/c mice. Spleen cells from such immunized mice are used to produce anti-Id hybridomas secreting anti-Id mAbs. Further, the anti-Id mAbs can be coupled to a carrier such as keyhole limpet hemocyanin (KLH) and used to immunize additional BALB/c mice. Sera from these mice will contain anti-anti-Id antibodies that have the binding properties of the original mAb specific for an epitope of the above MORT-1 protein, analogs, fragments or derivatives.

The anti-Id mAbs thus have their own idiotypic epitopes, or "idiotopes" structurally similar to the epitope being evaluated, such as GRB protein- α .

The term "antibody" is also meant to include both intact molecules as well as fragments thereof, such as, for example, Fab and F(ab')₂, which are capable of binding antigen. Fab and F(ab')₂ fragments lack the Fc fragment of intact antibody, clear more rapidly from the circulation, and may have less non-specific tissue binding than an intact antibody (Wahl et al., *J. Nucl. Med.* 24:316-325 (1983)).

It will be appreciated that Fab and F(ab')₂ and other fragments of the antibodies useful in the present invention may be used for the detection and quantitation of the MORT-1 protein according to the methods disclosed herein for intact antibody molecules. Such fragments are typically produced by proteolytic cleavage, using enzymes such as papain (to produce Fab fragments) or pepsin (to produce F(ab')₂ fragments).

An antibody is said to be "capable of binding" a molecule if it is capable of specifically reacting with the molecule to thereby bind the molecule to the antibody. The term "epitope" is meant to refer to that portion of any molecule capable of being bound by an antibody which can also be recognized by that antibody. Epitopes or "antigenic determinants" usually consist of chemically active surface groupings of molecules such as amino acids or sugar side chains and have specific three dimensional structural characteristics as well as specific charge characteristics.

An "antigen" is a molecule or a portion of a molecule capable of being bound by an antibody which is additionally capable of inducing an animal to produce antibody capable of binding to an epitope of that antigen. An antigen may have one or more than one epitope. The specific reaction referred to above is meant to indicate that the antigen will react, in a highly selective manner, with its corresponding antibody and not with the multitude of other antibodies which may be evoked by other antigens.

The antibodies, including fragments of antibodies, useful in the present invention may be used to quantitatively or qualitatively detect the MORT-1 or MORT-1-binding protein in a sample or to detect presence of cells which express the MORT-1 or MORT-1-binding protein of the present invention. This can be accomplished by immunofluorescence techniques employing a fluorescently labeled antibody (see below) coupled with light microscopic, flow cytometric, or fluorometric detection.

The antibodies (or fragments thereof) useful in the present invention may be employed histologically, as in immunofluorescence or immunoelectron microscopy, for *in situ* detection of the MORT-1 or MORT-1-binding protein of the present invention. *In situ* detection may be accomplished by removing a histological specimen from a patient, and providing the labeled antibody of the present invention to such a specimen. The antibody (or fragment) is preferably provided by applying or by overlaying the labeled antibody (or fragment) to a biological sample. Through the use of such a procedure, it is possible to determine not only the presence of the MORT-1 or MORT-1-binding protein, but also its distribution on the examined tissue. Using the present invention, those of ordinary skill will readily perceive that any of wide variety of histological methods (such as staining procedures) can be modified in order to achieve such *in situ* detection.

Such assays for the MORT-1 or MORT-1-binding protein of the present invention typically comprises incubating a biological sample, such as a biological fluid, a tissue extract, freshly harvested cells such as lymphocytes or leukocytes, or cells which have been incubated in tissue culture, in the presence of a detectably labeled antibody capable of identifying the MORT-1 protein, and detecting the antibody by any of a number of techniques well known in the art.

The biological sample may be treated with a solid phase support or carrier such as nitrocellulose, or other solid support or carrier which is capable of immobilizing cells, cell particles or soluble proteins. The support or carrier may then be washed with suitable buffers followed by treatment with a detectably labeled antibody in accordance with the present invention, as noted above. The solid phase support or carrier may then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on said solid support or carrier may then be detected by conventional means.

By "solid phase support", "solid phase carrier", "solid support", "solid carrier", "support" or "carrier" is intended any support or carrier capable of binding antigen or antibodies. Well-known supports or carriers, include glass, polystyrene, polypropylene, polyethylene, dextran, nylon amyloses, natural and modified celluloses, polyacrylamides, gabbros and magnetite. The nature of the carrier can be either soluble to some extent or

insoluble for the purposes of the present invention. The support material may have virtually any possible structural configuration so long as the coupled molecule is capable of binding to an antigen or antibody. Thus, the support or carrier configuration may be spherical, as in a bead, cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface may be flat such as a sheet, test strip, etc. Preferred supports or carriers include polystyrene beads. Those skilled in the art will know many other suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

The binding activity of a given lot of antibody, of the invention as noted above, may be determined according to well known methods. Those skilled in the art will be able to determine operative and optimal assay conditions for each determination by employing routine experimentation.

Other such steps as washing, stirring, shaking, filtering and the like may be added to the assays as is customary or necessary for the particular situation.

One of the ways in which an antibody in accordance with the present invention can be detectably labeled is by linking the same to an enzyme and use in an enzyme immunoassay (EIA). This enzyme, in turn, when later exposed to an appropriate substrate, will react with the substrate in such a manner as to produce a chemical moiety which can be detected, for example, by spectrophotometric, fluorometric or by visual means. Enzymes which can be used detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods which employ a chromogenic substrate for the enzyme. Detection may also be accomplished by visual comparison of the extent of enzymatic reaction of a substrate in comparison with similarly prepared standards.

Detection may be accomplished using any of a variety of other immunoassays. For example, by radioactivity labeling the antibodies or antibody fragments, it is possible to detect R-PTPase through the use of a radioimmunoassay (RIA). A good description of RIA may be found in Laboratory Techniques and Biochemistry in Molecular Biology, by Work, T.S. et al., North Holland Publishing Company, NY (1978) with particular reference to the chapter entitled "An Introduction to Radioimmune Assay and Related Techniques" by Chard, T., incorporated by reference herein. The radioactive isotope can be detected by such means as the use of a γ counter or a scintillation counter or by autoradiography.

It is also possible to label an antibody in accordance with the present invention with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wavelength, its presence can be then detected due to fluorescence. Among the most commonly used fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthalaldehyde and fluorescamine.

The antibody can also be detectably labeled using fluorescence emitting metals such as ^{152}E , or others of the lanthanide series. These metals can be attached to the antibody using such metal chelating groups as diethylenetriamine pentaacetic acid (ETPA).

The antibody can also be detectably labeled by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction. Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, theromatic acridinium ester, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound may be used to label the antibody of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase and aequorin.

An antibody molecule of the present invention may be adapted for utilization in an immunometric assay, also known as a "two-site" or "sandwich" assay. In a typical

immunometric assay, a quantity of unlabeled antibody (or fragment of antibody) is bound to a solid support or carrier and a quantity of detectably labeled soluble antibody is added to permit detection and/or quantitation of the ternary complex formed between solid-phase antibody, antigen, and labeled antibody.

Typical, and preferred, immunometric assays include "forward" assays in which the antibody bound to the solid phase is first contacted with the sample being tested to extract the antigen from the sample by formation of a binary solid phase antibody-antigen complex. After a suitable incubation period, the solid support or carrier is washed to remove the residue of the fluid sample, including unreacted antigen, if any, and the contacted with the solution containing an unknown quantity of labeled antibody (which functions as a "reporter molecule"). After a second incubation period to permit the labeled antibody to complex with the antigen bound to the solid support or carrier through the unlabeled antibody, the solid support or carrier is washed a second time to remove the unreacted labeled antibody.

In another type of "sandwich" assay, which may also be useful with the antigens of the present invention, the so-called "simultaneous" and "reverse" assays are used. A simultaneous assay involves a single incubation step as the antibody bound to the solid support or carrier and labeled antibody are both added to the sample being tested at the same time. After the incubation is completed, the solid support or carrier is washed to remove the residue of fluid sample and uncomplexed labeled antibody. The presence of labeled antibody associated with the solid support or carrier is then determined as it would be in a conventional "forward" sandwich assay.

~~In the "reverse" assay, stepwise addition first of a solution of labeled antibody to the~~
fluid sample followed by the addition of unlabeled antibody bound to a solid support or carrier after a suitable incubation period is utilized. After a second incubation, the solid phase is washed in conventional fashion to free it of the residue of the sample being tested and the solution of unreacted labeled antibody. The determination of labeled antibody associated with a solid support or carrier is then determined as in the "simultaneous" and "forward" assays.

The MORT-1 and MORT-1-binding proteins of the invention may then be produced by any standard recombinant DNA procedure (see for example, Sambrook, et al., 1989) in which suitable eukaryotic or prokaryotic host cells are transformed by appropriate eukaryotic or prokaryotic vectors containing the sequences encoding for the proteins. Accordingly, the present invention also concerns such expression vectors and transformed hosts for the production of the proteins of the invention. As mentioned above, these proteins also include their biologically active analogs, fragments and derivatives, and thus the vectors encoding them also include vectors encoding analogs and fragments of these proteins, and the transformed hosts include those producing such analogs and fragments. The derivatives of these proteins are the derivatives produced by standard modification of the proteins or their analogs or fragments, produced by the transformed hosts.

The present invention also relates to pharmaceutical compositions comprising recombinant animal virus vectors encoding the MORT-1 protein or MORT-1-binding proteins, which vector also encodes a virus surface protein capable of binding specific target cell (e.g. cancer cells) surface proteins to direct the insertion of the MORT-1 or MORT-1-binding protein sequences into the cells. Other aspects of the invention will be apparent from the following examples.

The invention will now be described in more detail in the following non-limiting examples and the accompanying drawings :

EXAMPLE 1: Cloning and isolation of the MORT-1 protein which binds to the intracellular domain of the FAS-R

(i) Two-hybrid screen and two-hybrid β -galactosidase expression test

To isolate proteins interacting with the intracellular domain of the FAS-R, the yeast two-hybrid system was used (Fields and Song, 1989; see also co-pending IL 109632, 112002

and 112692). Briefly, this two-hybrid system is a yeast-based genetic assay to detect specific protein-protein interactions *in vivo* by restoration of a eukaryotic transcriptional activator such as GAL4 that has two separate domains, a DNA binding and an activation domain, which domains when expressed and bound together to form a restored GAL4 protein, is capable of binding to an upstream activating sequence which in turn activates a promoter that controls the expression of a reporter gene, such as lacZ or HIS3, the expression of which is readily observed in the cultured cells. In this system the genes for the candidate interacting proteins are cloned into separate expression vectors. In one expression vector the sequence of the one candidate protein is cloned in phase with the sequence of the GAL4 DNA-binding domain to generate a hybrid protein with the GAL4 DNA-binding domain, and in the other vector the sequence of the second candidate protein is cloned in phase with the sequence of the GAL4 activation domain to generate a hybrid protein with the GAL4-activation domain. The two hybrid vectors are then co-transformed into a yeast host strain having a lacZ or HIS3 reporter gene under the control of upstream GAL4 binding sites. Only those transformed host cells (cotransformants) in which the two hybrid proteins are expressed and are capable of interacting with each other, will be capable of expression of the reporter gene. In the case of the lacZ reporter gene, host cells expressing this gene will become blue in color when X-gal is added to the cultures. Hence, blue colonies are indicative of the fact that the two cloned candidate proteins are capable of interacting with each other.

Using this two-hybrid system, the intracellular domain, FAS-IC, was cloned, separately, into the vector pGBT9 (carrying the GAL4 DNA-binding sequence, provided by CLONTECH, USA, see below), to create fusion proteins with the GAL4 DNA-binding domain. For the cloning of FAS-R into pGBT9, a clone encoding the full-length cDNA sequence of FAS-R (see co-pending IL 111125) was used from which the intracellular domain (IC) was excised by standard procedures using various restriction enzymes and then isolated by standard procedures and inserted into the pGBT9 vector opened, in its multiple cloning site region (MCS), with the corresponding suitable restriction enzymes. It should be noted that the FAS-IC extends from residues 175-319 of the intact FAS-R, this portion

containing residues 175-319 being the FAS-IC inserted into the pGBT9 vector (see also IL 111125).

The above hybrid (chimeric) vector was then cotransfected together with a cDNA library from human HeLa cells cloned into the pGAD GH vector, bearing the GAL4 activating domain, into the HF7c yeast host strain (all the above-noted vectors, pGBT9 and pGAD GH carrying the HeLa cell cDNA library, and the yeast strain were purchased from Clontech Laboratories, Inc., USA, as a part of MATCHMAKER Two-Hybrid System, #PT1265-1). The co-transfected yeasts were selected for their ability to grow in medium lacking Histidine (His⁻ medium), growing colonies being indicative of positive transformants. The selected yeast clones were then tested for their ability to express the lacZ gene, i.e. for their LAC Z activity, and this by adding X-gal to the culture medium, which is catabolized to form a blue colored product by β -galactosidase, the enzyme encoded by the lacZ gene. Thus, blue colonies are indicative of an active lacZ gene. For activity of the lacZ gene, it is necessary that the GAL4 transcription activator be present in an active form in the transformed clones, namely that the GAL4 DNA-binding domain encoded by the above hybrid vector be combined properly with the GAL4 activation domain encoded by the other hybrid vector. Such a combination is only possible if the two proteins fused to each of the GAL4 domains are capable of stably interacting (binding) to each other. Thus, the His⁺ and blue (LAC Z⁺) colonies that were isolated are colonies which have been cotransfected with a vector encoding FAS-IC and a vector encoding a protein product of human HeLa cell origin that is capable of binding stably to FAS-IC.

The plasmid DNA from the above His⁺, LAC Z⁺ yeast colonies was isolated and electroporated into E. coli strain HB101 by standard procedures followed by selection of Leu⁺ and Ampicillin resistant transformants, these transformants being the ones carrying the hybrid pGAD GH vector which has both the Amp^R and Leu² coding sequences. Such transformants therefore are clones carrying the sequences encoding newly identified proteins capable of binding to the FAS-IC. Plasmid DNA was then isolated from these transformed E. coli and retested by :

(a) retransforming them with the original FAS-R intracellular domain hybrid plasmid (hybrid pGTB9 carrying the FAS-IC) into yeast strain HF7 as set forth hereinabove. As controls, vectors carrying irrelevant protein encoding sequences, e.g. pACT-lamin or pGBT9 alone were used for cotransformation with the FAS-IC-binding protein (i.e. MORT-1)-encoding plasmid. The cotransformed yeasts were then tested for growth on His⁻ medium alone, or with different levels of 3-aminotriazole; and

(b) retransforming the plasmid DNA and original FAS-IC hybrid plasmid and control plasmids described in (a) into yeast host cells of strain SFY526 and determining the LAC Z⁺ activity (effectivity of β -gal formation, i.e. blue color formation).

The results of the above tests revealed that the pattern of growth of colonies in His⁻ medium was identical to the pattern of LAC Z activity, as assessed by the color of the colony, i.e. His⁺ colonies were also LAC Z⁺. Further, the LAC Z activity in liquid culture (preferred culture conditions) was assessed after transfection of the GAL4 DNA-binding and activation-domain hybrids into the SFY526 yeast hosts which have a better LAC Z inducibility with the GAL4 transcription activator than that of the HF-7 yeast host cells.

Using the above procedure a protein called HF1, and now also called MORT-1 for "Mediator of Receptor-induced Toxicity", was identified, isolated and characterized.

Furthermore, it should also be mentioned that in a number of the above two-hybrid β -galactosidase expression tests, the expression of β -galactosidase was also assessed by a preferred filter assay. In the screening, 5 of about 3×10^6 cDNAs were found to contain the MORT-1 insert. The so-isolated cloned MORT-1 cDNA inserts were then sequenced using standard DNA sequencing procedures. The amino acid sequence of MORT-1 was deduced

from the DNA sequence. Residue numbering in the proteins encoded by the cDNA inserts are as in the Swiss-Prot data bank. Deletion mutants were produced by PCR, and point mutants by oligonucleotide-directed mutagenesis (Current Protocols in Molec. Biol., 1994).

(ii) Induced expression, metabolic labeling and immunoprecipitation of proteins

MORT-1, N-linked to the FLAG octopeptide (FLAG-HF1; Eastman Kodak, New Haven, Ct., USA), Fas-IC, FAS-R, p55-R, a chimera comprised of the extracellular domain of p55-R (amino acids 1-168) fused to the transmembrane and intracellular domain of FAS-R

(amino acids 153-319), and the luciferase cDNA which serves as a control, were expressed in HeLa cells. Expression was carried out using a tetracycline-controlled expression vector, in a HeLa cell clone (HtTA-1) that expresses a tetracycline-controlled transactivator (Gossen and Bujard, 1992; as described in IL 109632, 111125, 112002 and 112692; see also Boldin et al., 1995). Metabolic labeling with [35 S]methionine and [35 S]cysteine (DUPONT, Wilmington, De., USA and Amersham, Buckinghamshire, England) was performed 18 hours after transfection, by a further 4h incubation at 37°C in Dulbecco's modified Eagle's medium lacking methionine and cysteine, but supplemented with 2% dialyzed fetal calf serum. The cells were then lysed in RIPA buffer (10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1% NP-40, 1% deoxycholate, 0.1% SDS and 1 mM EDTA) and the lysate was precleared by incubation with irrelevant rabbit antiserum (3 μ l/ml) and Protein G Sepharose beads (Pharmacia, Uppsala, Sweden; 60 μ l/ml). Immunoprecipitation was performed by 1h incubation at 4°C of 0.3 ml aliquots of lysate with mouse monoclonal antibodies (5 μ l/aliquot) against the FLAG octopeptide (M2; Eastman Kodak), p55-R (#18 and #20; (Engelmann et al., 1990)), or FAS-R (ZB4; Kamiya Southand Oaks, Ca., USA), or with isotype matched mouse antibodies as a control, followed by a further 1h incubation with Protein G Sepharose beads (30 μ l/aliquot).

(iii) *In vitro* binding

Glutathione S-transferase (GST) fusions with the wild type or a mutated Fas-IC were produced and adsorbed to glutathione-agarose beads (as described in IL 109632, 111125, 112002, 112692; see also Boldin et al., 1995; Current protocols in molecular biology, 1994; Frangioni and Neel, 1993). Binding of metabolically-labeled FLAG-HF1 fusion protein to GST-Fas-IC was assessed by incubating the beads for 2h at 4°C with extracts of HeLa cells, metabolically labeled with [35 S]methionine (60 μ Ci/ml), that express FLAG-HF1. The extracts were prepared in a buffer containing 50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 0.1% NP-40, 1 mM dithiotreitol, 1 mM EDTA, 1 mM phenylmethylsulfonyl fluoride, 20 μ g/ml Aprotinin, 20 μ g/ml Leupeptin, 10 mM sodium fluoride and 0.1 mM sodium vanadate (1 ml per 5×10^5 cells).

(iv) Assessment of the cytotoxicity triggered by induced expression of HF1

HF1, Fas-IC, p55-IC and luciferase cDNAs were inserted into a tetracycline-controlled expression vector and transfected to HtTA-1 cells (a HeLa cell line) (Gossen and Bujard, 1992) together with the secreted placental alkaline phosphatase cDNA, placed under control of SV40 promoter (the pSBC-2 vector (Dirks et al., 1993)). Cell death was assessed 40 hours after transfection, either by the neutral-red uptake assay (Wallach, 1984) or, for assessing specifically the death in those cells that express the transfected cDNAs, by determining the amounts of placental alkaline phosphatase (Berger et al., 1988) secreted to the growth medium at the last 5 hours of incubation.

In another set of experiments to analyze the region of the MORT-1 (HF1) protein involved in the binding to the FAS-IC, the following proteins were expressed transiently in HeLa cells that contain a tetracycline-controlled transactivator (HtTA-1), using a tetracycline-controlled expression vector (pUHD10-3): Human FAS-R alone; Human FAS-R as well as the N-terminal part of MORT-1 (amino acids 1-117, the "MORT-1 head"); Human FAS-R as well as the C-terminal part of MORT-1, which contains its 'death domain' homology region (amino acids 130-245, the "MORT-1 DD", see also IL 112742); FLAG-55.11 (amino acids 309-900 of protein 55.11 fused at the N-terminus to the FLAG octapeptide, the protein 55.11 being a p55-IC-specific binding protein, see also IL 109632). 12 hours after transfection, the cells were trypsinized and re-seeded at a concentration of 30,000 cells/well. After 24 hrs further incubation, the cells were treated for 6 hrs with a monoclonal antibody against the extracellular domain of FAS-R (monoclonal antibody CH-11, Oncor) at various concentrations (0.001-10 µg/ml monoclonal antibody), in the presence of 10 g/ml cycloheximide. Cell viability was then determined by the neutral-red uptake assay and the results were presented in terms of % viable cells as compared to cells that had been incubated with cycloheximide alone (in the absence of anti-FAS-R monoclonal antibody CH-11).

(v) Northern and sequence analyses

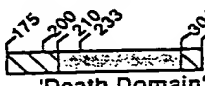
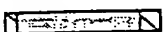
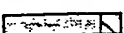
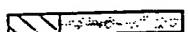
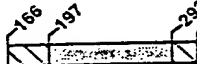
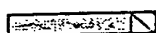


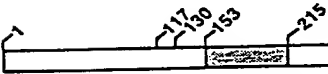

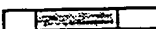
poly A⁺ RNA was isolated from total RNA of HeLa cells (Oligotex-dT mRNA kit. QIAGEN, Hilden, Germany). Northern analysis using the HF1 cDNA as a probe was

performed by conventional methods (as described in IL 109632; see also Boldin et al., 1995). The nucleotide sequence of MORT-1 (HF1) was determined in both directions by the dideoxy chain termination method.

The results (shown in Table 1 and Figs. 1-7) obtained from the above experimental procedures are as follows: Sequence analysis of the MORT-1 cDNA cloned by the two-hybrid procedure indicated that it encodes a novel protein (see below). Applying the two-hybrid test further to evaluate the specificity of the binding of this protein (MORT-1) for "Mediator of Receptor-induced Toxicity") to Fas-IC, and to define the particular region in Fas-IC to which it binds, led to the following findings (Table 1): (a) The protein binds both to human and to mouse Fas-IC, but not to several other tested proteins, including three receptors of the TNF/NGF receptor family (p55 and p75 TNF receptors and CD40); (b) Replacement mutations at position 225 (Ile) in the 'death domain' of FAS-R, shown to abolish signaling both *in vitro* and *in vivo* (the *lpr^{CG}* mutation (Watanabe-Fukunaga et al., 1992; Itoh and Nagata, 1993), also prevents binding of MORT-1 to the FAS-IC; (c) The MORT-1 binding-site in FAS-R occurs within the 'death domain' of this receptor; and (d) MORT-1 binds to itself. This self-binding, and the binding of HF1 to FAS-R involve different regions of the protein: A fragment of MORT-1 corresponding to residues 1-117 binds to the full-length MORT-1, but does not bind to itself nor to the FAS-IC. Conversely, a fragment corresponding to residues 130-245 binds to FAS-R, yet does not bind to MORT-1 (Table 1). Furthermore, it is apparent from the results in Table 1 that the 'death domain' region of FAS-R is critical for FAS-IC self-association, as is the 'death domain' region of p55-R for p55-IC self-association. The deletions on both sides of these 'death domains' does not affect the self-association ability thereof while, however, a deletion within these 'death domains' does affect the self-association. In the case of MORT-1, the binding of MORT-1 to FAS-IC is also dependent upon the complete (full) 'death domain' of FAS-R, while however, it is also not dependent on the regions outside of the FAS-R 'death domain' region for FAS-IC binding.

Table 1

Interaction of MORT-1 with FAS-IC and self-association of MORT-1 within transformed yeasts: assessment by a two-hybrid β -galactosidase expression test

DNA-BINDING - DOMAIN HYBRID	ACTIVATION-DOMAIN HYBRID					
	MORT1			human Fas-IC	SNF4	PGAD-GH
	Full	1-117	130-245			
Human Fas-IC						
 Full	++	-	+	++	-	-
 200-319	++			++	-	-
 233-319	-			-	-	-
 175-304	++	-	+	++	-	-
Mouse Fas-IC						
 Full	++	-	++	++	-	-
 197-306	++	-	+	++	-	-
 I225N	-	-	-	++	-	-
 I225A	-			++	-	-
MORT1						
 Full	++	++	-	++	-	-
 1-117	+	-		-	-	-
 130-245	-		-	++	-	-

SPECIFICITY TESTS

human p55-IC	-	-	-	-
human p75-IC	-	-	-	-
human CD40-IC	-	-	-	-
Cyclin D	-	-	-	-
Lamin	-	-	-	-
SNF1	-	-	+	-
pGBT9	-	-	-	-

In Table 1 above there is depicted the interaction of the proteins encoded by the Gal4 DNA binding domain and activation-domain constructs (pGBT9 and pGAD-GH) within transfected SFY526 yeasts as assessed by β -galactosidase expression filter assay. The DNA-binding-domain constructs included four constructs of the human Fas-IC, four constructs of the mouse Fas-IC including two full-length constructs having Ile to Leu or Ile to Ala replacement mutations at position 225 (I225N and I225A, respectively), and three HF1 (MORT-1) constructs, all of which constructs are shown schematically on the left-hand side of the table. The activation-domain constructs included three HF1 constructs, the HF1 portion being as in the DNA-binding-domain constructs; and a full-length human Fas-IC construct, the Fas-IC portion being the same as in the above DNA-binding domain construct. The intracellular domains of human p55 TNF receptor (p55-IC residues 206-426), human CD40 (CD40-IC, residues 216-277) and human p75 TNF receptor (p75-IC, residues 287-461) as well as lamin, cyclin D and 'empty' Gal4 (pGBT9) vectors served as negative controls in the form of DNA-binding domain constructs. SNF-1 and SNF4 served as positive controls in the form of DNA-binding-domain (SNF1) and activation-domain (SNF4) constructs. 'Empty' Gal4 vectors (pGAD-GH) also served as negative controls in the form of activation domain constructs (for more details concerning the p55-IC, p75-IC, see also IL 109632, 111125, 112002, 112692). The symbols "++" and "+" denote the development of strong color within 30 and 90 min of the assay, respectively; and "-" denotes no development of color within 24h. Combinations for which no score is given have not been tested.

Expression of HF1 (MORT-1) molecules fused at their N terminus with the FLAG octapeptide (FLAG-HF1) yielded in HeLa cells proteins of four distinct sizes—about 27, 28, 32, and 34 kD. In Fig. 1 (A and B) there is shown the results demonstrating the interaction of HF1 with Fas-IC *in vitro*. As noted above in the description of Figs. 1A and B, Fig. 1A is a reproduction of a control autoradiogram of an immunoprecipitate of proteins from extracts of HeLa cells transfected with the FLAG-HF1 (FLAG-MORT1) fusion protein or with luciferase cDNA as a control, the immunoprecipitation being performed with anti-FLAG antibody (α FLAG). Fig. 1B is a reproduction of an autoradiogram showing the interaction *in vitro* between HF1 and FAS-IC wherein the HF1 is in the form of [35 S]methionine-

metabolically labeled HF1-FLAG fusion proteins obtained from extracted of transfected HeLa cells and the FAS-IC is in the form of human and mouse GST-FAS-IC fusion proteins including one having a replacement mutation at position 225 in FAS-IC, all of which GST-FAS-IC fusion proteins were produced in *E. coli*. The GST-fusion proteins were attached to glutathione beads before interaction with the extracts containing the HF1-FLAG fusion protein following this interaction, SDS-PAGE was performed. Thus the *in vitro* interaction was evaluated by assessing, by autoradiography following SDS-PAGE, the binding of [³⁵S] metabolically labeled HF1, produced in transfected HeLa cells as a fusion with the FLAG octapeptide (FLAG-HF1), to GST, GST fusion with the human or mouse Fas-IC (GST-huFas-IC, GST-mFas-IC) or to GST fusion with Fas-IC containing a Ile to Ala replacement mutation at position 225. As is apparent from Fig. 1B, all four FLAG-HF1 proteins showed ability to bind to Fas-IC upon incubation with a GST-Fas-IC fusion protein. As in the yeast two-hybrid test (Table 1), HF1 did not bind to a GST-Fas-IC fusion protein with a replacement at the *lpr^{CG}* mutation site (I225A).

The proteins encoded by the FLAG-HF1 cDNA showed also an ability to bind to the intracellular domain of FAS-R, as well as to the intracellular domain of FAS-R chimera whose extracellular domain was replaced with that of p55-R (p55-FAS), when co-expressed with these receptors in HeLa cells. In Fig. 2 (A, B, C) there is shown the results demonstrating the interaction of HF1 with FAS-IC in transfected HeLa cells, i.e. *in vivo*. As mentioned above in the description of Figs. 2 A, B, C, these figures are reproductions of autoradiograms of immunoprecipitates of various transfected HeLa cells which demonstrate the *in vivo* interaction and specificity of the interaction between HF1 and FAS-IC in cells co-

transfected with constructs encoding these proteins. Thus, FLAG-HF1 fusion protein was expressed and metabolically labeled with [³⁵S] cystein (20 µCi/ml) and [³⁵S]methionine (40 µCi/ml) in HeLa cells, alone, or together with human FAS-R, FAS-R chimera in which the extracellular domain of FAS-R was replaced with the corresponding region in the human p55-R (p55-FAS), or the human p55-R, as negative control. Cross immunoprecipitation of HF1 with the co-expressed receptor was performed using the indicated antibodies (Figs. 2 A-C). As is apparent in Figs. 2 A-C, FLAG-HF1 is capable of binding to the intracellular

domain of FAS-R, as well as to the intracellular domain of a FAS-R-p55-R chimera having the extracellular domain of p55-R and the intracellular domain of FAS-R, when co-expressed with these receptors in the HeLa cells (see 3 middle lanes Fig. 2A and 3 left-hand lanes Fig. 2C, respectively). Further, immunoprecipitation of FLAG-HF1 from extracts of the transfected cells also resulted in precipitation of the co-expressed FAS-R (Fig. 2A) or the co-expressed p55-FAS chimera (Fig. 2C). Conversely, immunoprecipitation of these receptors resulted in the coprecipitation of the FLAG-HF1 (Figs. 2A and 2C).

Northern analysis using the HF1 cDNA as probe revealed a single hybridizing transcript in HeLa cells. Fig. 3 shows a reproduction of a Northern blot in which poly A⁺ RNA (0.3 µg) from transfected cells was hybridized with the HF1 cDNA. The size of this transcript (about 1.8 kB) is close to that of the HF1 cDNA (about 1702 nucleotides).

In sequence analysis, the cDNA was found to contain an open reading frame of about 250 amino acids. Fig. 4 depicts the preliminary nucleotide and deduced amino acid sequence of HF1 in which the 'death domain' motif is underlined, as is a possible start Met residue (position 49; bold, underlined M) and the translation stop codon (the asterisk under the codon at position 769-771). This 'death domain' motif shares homology with the known p55-R and FAS-R 'death domain' motifs (p55DD and FAS-DD). In order to determine the precise C-terminal end of HF1 and to obtain evidence concerning the precise N-terminal (initial Met residue) end of HF1, additional experiments were carried out as follows:

Using the methods described above, a number of constructs encoding HF1 molecules fused at their N-terminus with the FLAG octapeptide (FLAG-HF1) were constructed and expressed in HeLa cells with metabolic labeling of the expressed proteins using ³⁵S-cysteine and ³⁵S-methionine (see the above in respect of Fig. 5B). The HF1-FLAG molecules were encoded by the following cDNAs containing different portions of the HF1-encoding sequence:

- i) The FLAG octapeptide cDNA linked to the 5' end of the HF1 cDNA from which nucleotides 1-145 (see Fig. 4) have been deleted;
- ii) The FLAG octapeptide cDNA linked to the 5' end of the HF1 full length cDNA (see FLAG-HF1 construct above in respect of Fig. 1B);

iii) The FLAG octapeptide cDNA linked to the 5' end of the HF1 cDNA from which nucleotides 1-145 as well as nucleotides 832-1701 (see Fig. 4) had been deleted and the codon GCC at position 142-144 was mutated to TCC to prevent start of translation at this site.

Following expression of the above HF1-FLAG fusion products, immunoprecipitation was carried out as mentioned above, using either anti-FLAG monoclonal antibodies (M2) or as a control, anti-p75 TNF-R antibodies (#9), followed by SDS-PAGE (10% acrylamide) and autoradiography. The results are shown in Fig. 5 which is a reproduction of an autoradiogram on which was separated the above noted HF1-FLAG fusion proteins, the samples loaded on each lane of the gel being as follows:

Lanes 1 and 2: HF1-FLAG fusion protein encoded by the FLAG octapeptide cDNA linked to the 5' end of the HF1 cDNA from which nucleotides 1-145 had been deleted.

Lanes 3 and 4: HF1-FLAG fusion protein encoded by the FLAG octapeptide cDNA linked to the 5' end of the full length HF1 cDNA.

Lanes 5 and 6: HF1-FLAG fusion protein encoded by the FLAG octapeptide cDNA linked to the 5' end of the HF1 cDNA from which nucleotides 1-145 as well as 832-1701 had been deleted and the GCC at position 142-144 was mutated to TCC to prevent start of translation at this site.

The immunoprecipitations were with anti-FLAG monoclonal antibodies for the samples in lanes 2, 4 and 6 and anti-p75 TNF-R antibodies for the samples in lanes 1, 3 and 5.

From the autoradiogram of Fig. 5 it is apparent that the identity of product sizes in lanes 2 and 4 confirms that the nucleotides 769-771 are the site of translation termination for HF1, i.e. this codon represents a stop signal and is indicated by an asterisk in Fig. 4. Further, the occurrence of a broad band which represents just two translation products (as seen on the gel but being strongly labeled becomes a single large band on the autoradiogram) in lane 6 indicates that the occurrence of two additional products (the higher molecular weight broad bands) in lanes 2 and 4 reflect the initiation of translation both at the N-terminus of the FLAG-HF1 fusion molecule and at the methionine residue number 49 within the HF1 sequence (see bold underlined M at position 49 of the amino acid sequence in Fig. 4). Thus,

the above results have confirmed (validated) the C-terminal end of HF1 and have provided evidence that the N-terminal end of HF1 may be at position 49 of the sequence in Fig. 4.

Indeed, it has been shown by additional expression experiments of HF1 without the FLAG octapeptide fused to its 5-end, that Met⁴⁹ serves as an effective site of translation initiation.

It should be mentioned that a search conducted in the 'Gene Bank' and 'Protein Bank' DataBases revealed that there is no sequence corresponding to that of HF1 depicted in Fig. 4. Thus, HF1 represents a new FAS-IC-specific binding protein.

High expression of p55-IC results in triggering of a cytotoxic effect (see IL 109632, 111125 and Boldin et al., 1995). The expression of Fas-IC in HeLa cells also has such an effect, though to a lower extent, which could be detected only with the use of a sensitive assay. In Fig. 6 A and B there is depicted graphically the ligand independent triggering of cytotoxic effects in cells transfected with HF1, as well as human p55-IC and FAS-IC. The effect of transient expression of HF1, human Fas-IC, human p55-IC, or luciferase that served as a control, on the viability of HeLa cells was assessed using a tetracycline-controlled expression vector. Cell viability was evaluated 40 min after transfecting these cDNAs either in presence (open bars, Figs. 6A and B) or absence (closed bars, Fig. 6A and B) of tetracycline (1 µg/ml, to block expression), together with a cDNA encoding the secreted placental alkaline phosphatase. Cell viability was determined either by the neutral red uptake assay (Fig. 6A) or, for determining specifically the viability of those particular cells that express the transfected DNA, by measuring the amounts of placental alkaline phosphatase secreted to the growth medium (Fig. 6B).

Thus, it is apparent from Figs. 6 A and B that the expression of HF1 in HeLa cells resulted in significant cell death, greater than that caused by FAS-IC expression. These cytotoxic effects of all of p55-IC, FAS-IC and HF1 seem to be related to the 'death domain' regions, present in all of these proteins, which 'death domains' have a propensity to self-associate, and thereby possibly prompting the cytotoxic effects.

In view of the above mentioned characteristics of HF1 (MORT-1), namely, the specific association of HF1 with that particular region in FAS-R which is involved in cell

death induction, and the fact that even a slight change of structure in that region, which prevents signaling (the *lpr^{CG}* mutation) abolishes also the binding of HF1, indicates that this protein plays a role in the signaling or triggering of cell death. This notion is further supported by the observed ability of HF1 to trigger by itself a cytotoxic effect. Thus, HF1 (MORT-1) may function as (i) a modulator of the self-association of FAS-R by its own ability to bind to FAS-R as well as to itself, or (ii) serve as a docking site for additional proteins that are involved in the FAS-R signaling, i.e. HF1 may be a 'docking' protein and may therefore bind other receptors besides FAS-R, or (iii) constitutes part of a distinct signaling system that interacts with FAS-R signaling.

In order to further analyze the regions of MORT-1 (HF1) involved in FAS-IC binding and modulation of the FAS-R-mediated cellular effects (cytotoxicity), the above-mentioned experiments were carried out, using vectors encoding portions of MORT-1 (the 'MORT-1 head', amino acids 101-17 and the 'MORT-1 dd', amino acids 130-245) (separately), with a vector encoding the human FAS-R for co-transfections of HeLa cells. In these experiments the various proteins and combinations of proteins were expressed transiently in HeLa cells that contain a tetracycline-controlled transactivator (HtTA-1) by inserting the sequences encoding the proteins into a tetracycline-controlled expression vector pUHD10-3. Control transfections employed vectors encoding only the FAS-R and vectors encoding the FLAG-55.11 fusion protein (the 55.11 protein being a p55-IC-specific binding protein of which a portion containing amino acids 309-900 was fused (at its N-terminal) to the FLAG octapeptide).

~~Following the transfection and incubation periods (see (iv) above) the transfected~~
 cells were treated with various concentrations of an anti-FAS-R monoclonal antibody (CH-11) which binds specifically to the extracellular domain of FAS-R expressed by cells. This binding of anti-FAS-R antibody induces the aggregation of the FAS-R at the cell surface (much like the FAS-R ligand) and induces the intracellular signaling pathway mediated by the FAS-IC, resulting, ultimately, in cell death (FAS-R mediated cell cytotoxicity). The concentrations of the anti-FAS-R monoclonal antibody (CH-11) used were in the range of

0.01-10 µg/ml, usually concentrations such as 0.005; 0.05; 0.5 and 5 µg/ml. The cells were treated with the anti-FAS antibody in the presence of 10 µg/ml cycloheximide.

The results of the above analysis are set forth graphically in Fig. 7 which depicts the % viability of the transfected cells as a function of the concentration of anti-FAS-R monoclonal antibody (CH11) used to treat the cells, for each of the four different groups of transfected cells. These groups of transfected cells are denoted by different symbols as follows : (i) the open squares denote cells transfected only with the non-relevant (i.e. non-FAS-IC binding) control vector encoding the FLAG-55.11 fusion protein ("55.11", negative control); (ii) the closed squares denote cells co-transfected with vectors encoding the FAS-R and vectors encoding the C-terminal portion of MORT-1, amino acids 130-245, which contains the MORT-1 'death domain (dd) homology region ("fas+mort1dd"); (iii) the closed triangles denote cells transfected with only the vector encoding the FAS-R ("fas", positive control); and (iv) the open circles denote cells co-transfected with vectors encoding the FAS-R and vectors encoding the N-terminal portion of MORT-1, amino acids 1-117, the 'MORT-1 head' ("fas+mort1he").

From the results presented in Fig. 7, it is apparent that the expression of FAS-R in the transfected cells conveys an increased sensitivity to the cytotoxic effects of the anti-FAS-R antibodies (compare "fas" to "55.11"). Further, the co-expression of the region in MORT-1 that contains the 'death domain' homology region and FAS-R ("fas+mort1dd") strongly interferes with FAS-induced (i.e. FAS-R mediated) cell death as would be expected from the ability (see Table 1 above) of the MORT-1 'death domain' (DD) region to bind to the FAS-R 'death domain' (FAS-DD). Moreover, co-expression of the N-terminal part of MORT-1 and FAS-R ("fas+mort1he") does not interfere with FAS-R-mediated cell death and, if at all, somewhat enhances the cytotoxicity (i.e. slightly increased cell death).

Thus, the above results clearly indicate that the MORT-1 protein has two distinct regions as far as binding to the FAS-IC and mediation of the cell-cytotoxic activity of the FAS-IC are concerned.

These results therefore also provide a basis for the use of different parts (i.e. active fragments or analogs) of the MORT-1 protein for different pharmaceutical applications. For

example, the analogs or fragments or derivatives thereof of the MORT-1 protein which contain essentially only the C-terminal portion of MORT-1 inclusive of its 'death domain' region may be used for inhibiting FAS-R-mediated cytotoxic effects in FAS-R containing cells or tissues and thereby protect these cells or tissues from the deleterious effects of the FAS-R ligand in cases such as, for example, acute hepatitis. Alternatively, the analogs or fragments or derivatives thereof of the MORT-1 protein which contain essentially only the N-terminal portion of MORT-1 may be used for enhancing the FAS-R-mediated cytotoxic effects in FAS-R containing cells and tissues, thereby leading to the enhanced destruction of these cells or tissues when desired in cases such as, for example, tumor cells and autoreactive T and B cells. As detailed herein above, the above uses of the different regions of MORT-1 may be carried out using the various recombinant viruses (e.g. Vaccinia) to insert the MORT-1 region-encoding sequence into specific cells or tissues it is desired to treat.

Furthermore, it is also possible to prepare and use various other molecules such as, antibodies, peptides and organic molecules which have sequences or molecular structures corresponding to the above noted MORT-1 regions in order to achieve the same desired effects mediated by these MORT-1 regions.

Moreover, MORT-1 may be utilized to specifically identify, isolate and characterize other proteins which are capable of binding to MORT-1 (i.e. MORT-1-binding proteins); see Example 2.

EXAMPLE 2

Isolation of a MORT-1 binding protein

(i) Two-hybrid screen and two-hybrid β -galactosidase expression test

In a manner analogous to the procedure described in Example 1, using the intracellular domain of p55 TNF-R (p55 IC) and MORT-1 as baits, and screening a human B-cell library, two cDNA clones were obtained, which encode a protein product capable of binding to both MORT-1 and p55-IC. Both clones had identical nucleotide sequences at the 5' end as set forth in Fig. 8.

(ii) **Binding properties of the newly cloned cDNA, in two hybrid screens**

Using the above-mentioned yeast two-hybrid procedure a construct containing the new MORT-1-binding protein cDNA was used as a "prey" to which were added constructs of a number of "baits" in separate reactions, to determine the binding specificity of the MORT-1-binding protein encoded by this cDNA. These "baits" included constructs encoding MORT-1, portions of MORT-1 (MORT 'head', aa1-117, MORT 'tail', aa 130-245), the p55 IC (206-426 p55) or portion thereof (the 'death domain', 326-426 p55; and others upstream of the 'death domain' i.e. 206-326). The results are shown in Table 2.

Table 2

Bait	β -galactosidase expression data
MORT-1	+++
130-245 MORT-1	+
1-117 MORT-1	-
206-426 p55	+++
326-426 p55	+++
206-326 p55	-
206-308 p55	-
206-345 p55	-
p55 L351NI	-
Fas IC	-
233-319 Fas	-
p75 IC	-
CD40 IC	-
pGBT10	-
SNF1	-
Cycline D	-
Lamin	-

The above results of the two-hybrid β -galactosidase expression test of the binding of the clone to a large panel of baits confirmed that the protein encoded by this clone binds specifically to the death domains of both the p55 TNF-R and MORT-1.

In general the MORT-1 binding protein may be utilized directly to modulate or mediate the MORT-1 associated effects on cells, or, indirectly, to modulate or mediate the FAS-R ligand effect on cells when this effect is modulated or mediated by MORT-1. The same holds true with respect to other intracellular proteins or intracellular domains of transmembrane proteins, as specifically demonstrated for the p55 TNF-R herein.

MORT-1-binding proteins include those which bind specifically to the entire MORT-1 protein or those which bind to different regions of the MORT-1 protein, e.g. the above-noted N- and C-terminal regions of MORT-1. The MORT-1-binding proteins which bind specifically to such regions may be used to modulate the activity of these regions and hence the specific activity of MORT-1 as determined by these regions.

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CLAIMS

1. A DNA sequence encoding a herein designated MORT-1 (or HF1) protein, analogs or fragments thereof, all of which are capable of binding to or interacting with the intracellular domain of the FAS ligand receptor (FAS-IC).
 2. A DNA sequence according to claim 1, selected from the group consisting of:
 - (a) a cDNA sequence derived from the coding region of a native MORT-1 protein;
 - (b) DNA sequences capable of hybridization to a sequence of (a) under moderately stringent conditions and which encode a biologically active FAS-R intracellular domain-binding protein; and
 - (c) DNA sequences which are degenerate as a result of the genetic code to the DNA sequences defined in (a) and (b) and which encode a biologically active FAS-R intracellular domain-binding protein.
 3. A DNA sequence according to claim 1 or claim 2 encoding the protein MORT-1 comprising the sequence depicted in Fig. 4.
 4. A MORT-1 protein, analogs or fragments and derivatives of any thereof encoded by a sequence according to any one of claims 1-3, said protein, analogs, fragments and derivatives being capable of binding to or interacting with the intracellular domain of the FAS-R.
 5. The protein MORT-1 according to claim 4, having the deduced amino acid sequence depicted in Fig. 4.
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6. A vector comprising a DNA sequence according to any one of claims 1-3.
 7. A vector according to claim 6 which is capable of being expressed in a eukaryotic host cell.
 8. A vector according to claim 6 which is capable of being expressed in a prokaryotic host cell.

9. Transformed eukaryotic or prokaryotic host cells containing a vector according to any one of claims 6-8.
10. A method for producing a protein, analogs, fragments or derivatives thereof according to claim 4, comprising growing the transformed host cells according to claim 9 under conditions suitable for the expression of said protein, analogs, fragments or derivatives, effecting post-translational modifications of said protein as necessary for obtention of said protein, analogs, fragments or derivatives and isolating said expressed protein, analogs, fragments or derivatives.
11. Antibodies or active fragments or derivatives thereof, specific for the MORT-1 protein, analogs, fragments or derivatives according to claim 4.
12. A method for the modulation of the FAS-R ligand effect on cells carrying a FAS-R, comprising treating said cells with one or more MORT-1 proteins, analogs, fragments or derivatives according to claim 4, capable of binding to the intracellular domain and modulating the activity of said FAS-R, wherein said treating of said cells comprises introducing into said cells said one or more proteins, analogs, fragments or derivatives in a form suitable for intracellular introduction thereof, or introducing into said cells a DNA sequence encoding said one or more proteins, analogs, fragments or derivatives in the form of a suitable vector carrying said sequence, said vector being capable of effecting the insertion of said sequence into said cells in a way that said sequence is expressed in said cells.
- ~~13. A method for the modulation of the FAS-R ligand-effect on cells according to claim~~
12, comprising treating said cells with MORT-1, analogs, fragments or derivatives thereof, capable of binding to the intracellular domain and modifying the activity of FAS-R, wherein said treating of cells comprises introducing into said cells said MORT-1, analogs, fragments or derivatives in a form suitable for intracellular introduction thereof, or introducing into said cells a DNA sequence encoding said MORT-1, analogs, fragments or derivatives in the form of a suitable vector carrying said

sequence, said vector being capable of effecting the insertion of said sequence into said cells in a way that said sequence is expressed in said cells.

14. A method according to claim 12 wherein said treating of said cells is by transfection of said cells with a recombinant animal virus vector comprising the steps of :

(a) constructing a recombinant animal virus vector carrying a sequence encoding a viral surface protein (ligand) that is capable of binding to a specific cell surface receptor on the surface of a FAS-R- carrying cell and a second sequence encoding a protein selected from the MORT-1 protein, analogs, fragments and derivatives according to claim 4, that when expressed in said cells is capable of modulating the activity of said FAS-R; and

(b) infecting said cells with said vector of (a).

15. A method for modulating the FAS-R ligand effect on cells carrying a FAS-R comprising treating said cells with antibodies or active fragments or derivatives thereof, according to claim 11, said treating being by application of a suitable composition containing said antibodies, active fragments or derivatives thereof to said cells, wherein when the MORT-1 protein or portions thereof of said cells are exposed on the extracellular surface, said composition is formulated for extracellular application, and when said MORT-1 proteins are intracellular said composition is formulated for intracellular application.

16. A method for modulating the FAS-R ligand effect on cells carrying a FAS-R ~~comprising treating said cells with an oligonucleotide sequence encoding an antisense~~ sequence of at least part of the MORT-1 sequence according to any one of claims 1-3, said oligonucleotide sequence being capable of blocking the expression of the MORT-1 protein.

17. A method according to claim 16 wherein said oligonucleotide sequence is introduced to said cells via a virus of claim 14 wherein said second sequence of said virus encodes said oligonucleotide sequence.

18. A method for treating tumor cells or HIV-infected cells or other diseased cells, comprising:
- (a) constructing a recombinant animal virus vector carrying a sequence encoding a viral surface protein capable of binding to a specific tumor cell surface receptor or HIV-infected cell surface receptor or receptor carried by other diseased cells and a sequence encoding a protein selected from the MORT-1 protein, analogs, fragments and derivatives of claim 4, that when expressed in said tumor, HIV-infected, or other diseased cell is capable of killing said cell; and
 - (b) infecting said tumor or HIV-infected cells or other diseased cells with said vector of (a).
19. A method for modulating the FAS-R ligand effect on cells comprising applying the ribozyme procedure in which a vector encoding a ribozyme sequence capable of interacting with a cellular mRNA sequence encoding a MORT-1 protein according to claim 4, is introduced into said cells in a form that permits expression of said ribozyme sequence in said cells, and wherein when said ribozyme sequence is expressed in said cells it interacts with said cellular mRNA sequence and cleaves said mRNA sequence resulting in the inhibition of expression of said MORT-1 in said cells.
20. A method selected from the method according to any one of claims 12-19, wherein said MORT-1 protein or said MORT-1 encoding sequence comprises at least that part of the MORT-1 protein, which binds specifically to the FAS-IC, or at least that part of the ~~MORT-1 encoding sequence that encodes that part of the MORT-1 protein which~~ binds specifically to the FAS-IC.
21. A method for isolating and identifying proteins, factors or receptors capable of binding to the MORT-1 protein according to claim 4, comprising applying the procedure of affinity chromatography in which said MORT-1 protein is attached to the affinity chromatography matrix, said attached protein is brought into contact with a cell

extract; and proteins, factors or receptors from the cell extract which bound to said attached protein are then eluted, isolated and analyzed.

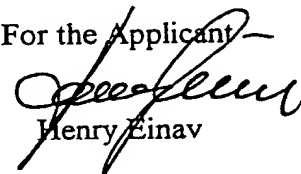
22. A method for isolating and identifying proteins, capable of binding to the MORT-1 protein according to claim 4, comprising applying the yeast two-hybrid procedure in which a sequence encoding said MORT-1 protein is carried by one hybrid vector and sequence from a cDNA or genomic DNA library is carried by the second hybrid vector, the vectors then being used to transform yeast host cells and the positive transformed cells being isolated, followed by extraction of the said second hybrid vector to obtain a sequence encoding a protein which binds to said MORT-1 protein.
 23. A pharmaceutical composition for the modulation of the FAS-R ligand- effect on cells comprising, as active, ingredient a MORT-1 protein according to claim 4, its biologically active fragments, analogs, derivatives or mixtures thereof.
 24. A pharmaceutical composition for modulating the FAS-R ligand- effect on cells comprising, as active ingredient, a recombinant animal virus vector encoding a protein capable of binding a cell surface receptor and encoding a MORT-1 protein or its biologically active fragments or analogs, according to claim 4.
 25. A pharmaceutical composition for modulating the FAS-R ligand effect on cells comprising as active ingredient, an oligonucleotide sequence encoding an anti-sense sequence of the MORT-1 sequence according to any one of claims 1-3.
 26. A method for isolating and identifying a protein capable of binding to the intracellular domain of FAS-R comprising applying the procedure of non-stringent southern hybridization followed by PCR cloning, in which a sequence or parts thereof according to any one of claims 1-3 is used as a probe to bind sequences from a cDNA or genomic DNA library, having at least partial homology thereto, said bound sequences then amplified and cloned by the PCR procedure to yield clones encoding proteins having at least partial homology to said sequences of claims 1-3.
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27. A method for the modulation of the MORT-1-induced effect on cells comprising treating said cells in accordance with a method according to any one of claims 12-19 with MORT-1, analogs, fragments or derivatives thereof or with sequences encoding MORT-1, analogs or fragments thereof, said treatment resulting in the enhancement or inhibition of said MORT-1-mediated effect.
28. A method according to claim 27 wherein said MORT-1 protein, analog, fragment or derivative thereof is that part of MORT-1 which is specifically involved in binding to MORT-1 itself, or said MORT-1 sequence encodes that part of MORT-1 which is specifically involved in binding to MORT-1 itself.
29. A DNA sequence encoding a protein capable of binding to MORT-1.
30. A DNA sequence according to claim 29 selected from the group consisting of :
 - (a) a cDNA sequence derived from the coding region of a native MORT-1-binding protein;
 - (b) DNA sequences capable of hybridization to a sequence of (a) under moderately stringent conditions and which encode a biologically active MORT-1-binding protein; and
 - (c) DNA sequences which are degenerate as a result of the genetic code to the DNA sequences defined in (a) and (b) and which encode a biologically active MORT-1-binding protein.
31. A MORT-1-binding protein or analogs, fragments and derivatives thereof encoded by a sequence according to claim 29 or claim 30.
- ~~32. A vector comprising a DNA sequence according to claim 29 or 30, which is capable of~~
being expressed in a prokaryotic or eukaryotic host cell.
33. Transformed eukaryotic or prokaryotic host cells containing a vector according to claim 32.
34. A method for producing the MORT-1-binding protein, analogs, fragments or derivatives thereof according to claim 31, comprising growing the transformed host cells according to claim 33 under conditions suitable for the expression of said protein, analogs or derivatives, effecting post-translational modifications of said protein as

necessary for obtention of said protein, analogs, fragments or derivatives and isolating said expressed protein, analogs, fragments or derivatives.

35. Antibodies or active fragments or derivatives thereof, specific for the MORT-1-binding protein, analogs, fragments or derivatives according to claim 31.
36. A method for the modulation of the MORT-1-associated effect on cells according to claim 27, said method being selected from the methods of any one of claims 12-19, wherein the MORT-1-binding protein, analogs or derivatives thereof or sequences encoding the MORT-1-binding protein, or analogs thereof are introduced into the cells and result in the inhibition or enhancement of the MORT-1-associated effect.

For the Applicant

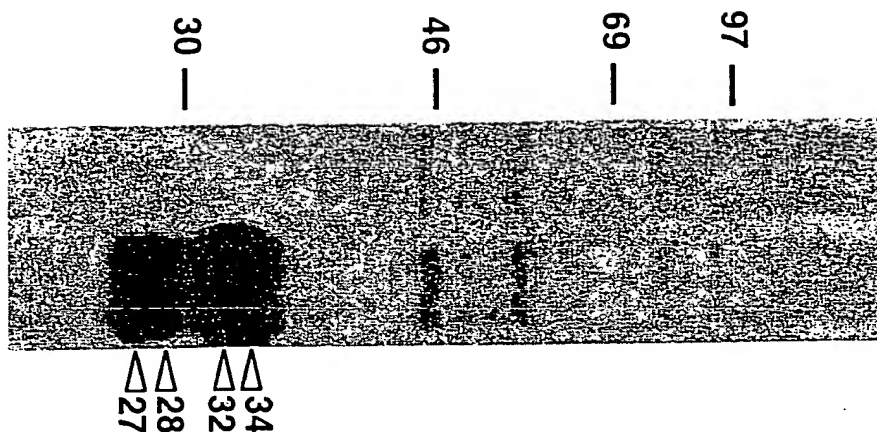


Henry Einav

**Immunoprecip.
with α FLAG Abs**

Proteins
expressed
in the
lysed cells:

LUCIFERASE
FLAG-MORT1



Binding to GST fusion proteins

GST	GST-huFas-IC	GST-mFas-IC	GST-mFas-IC I225A
LUCIFERASE	LUCIFERASE	LUCIFERASE	LUCIFERASE
FLAG-MORT1	FLAG-MORT1	FLAG-MORT1	FLAG-MORT1

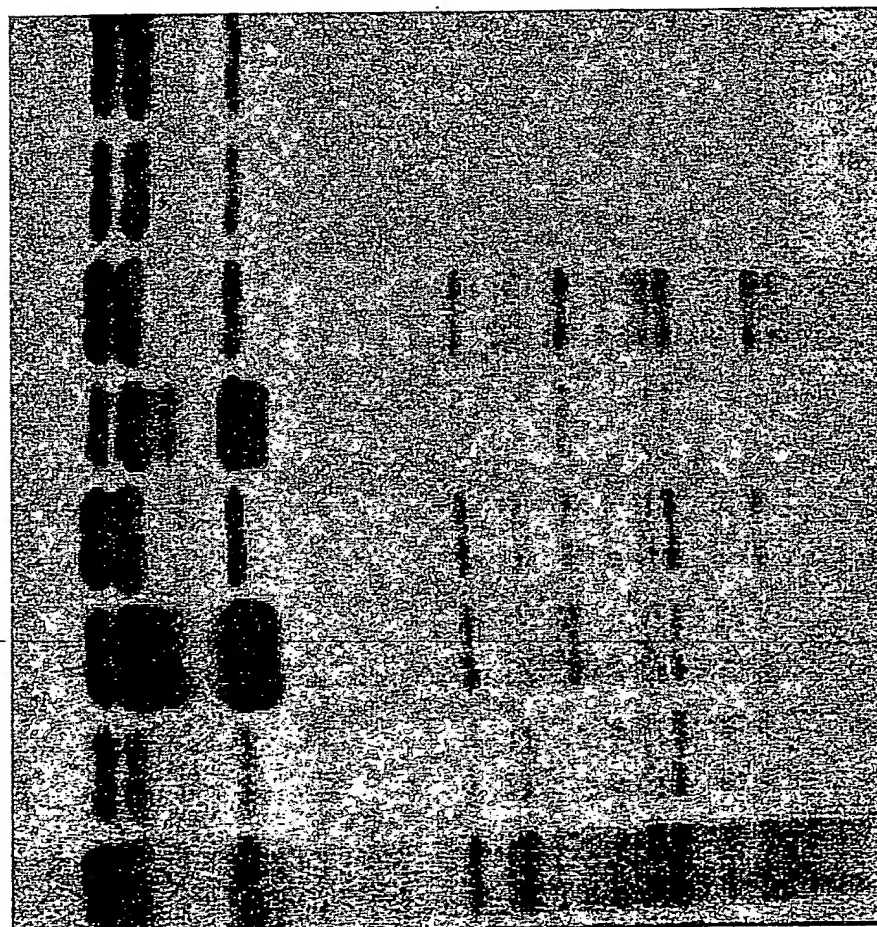


FIGURE 1

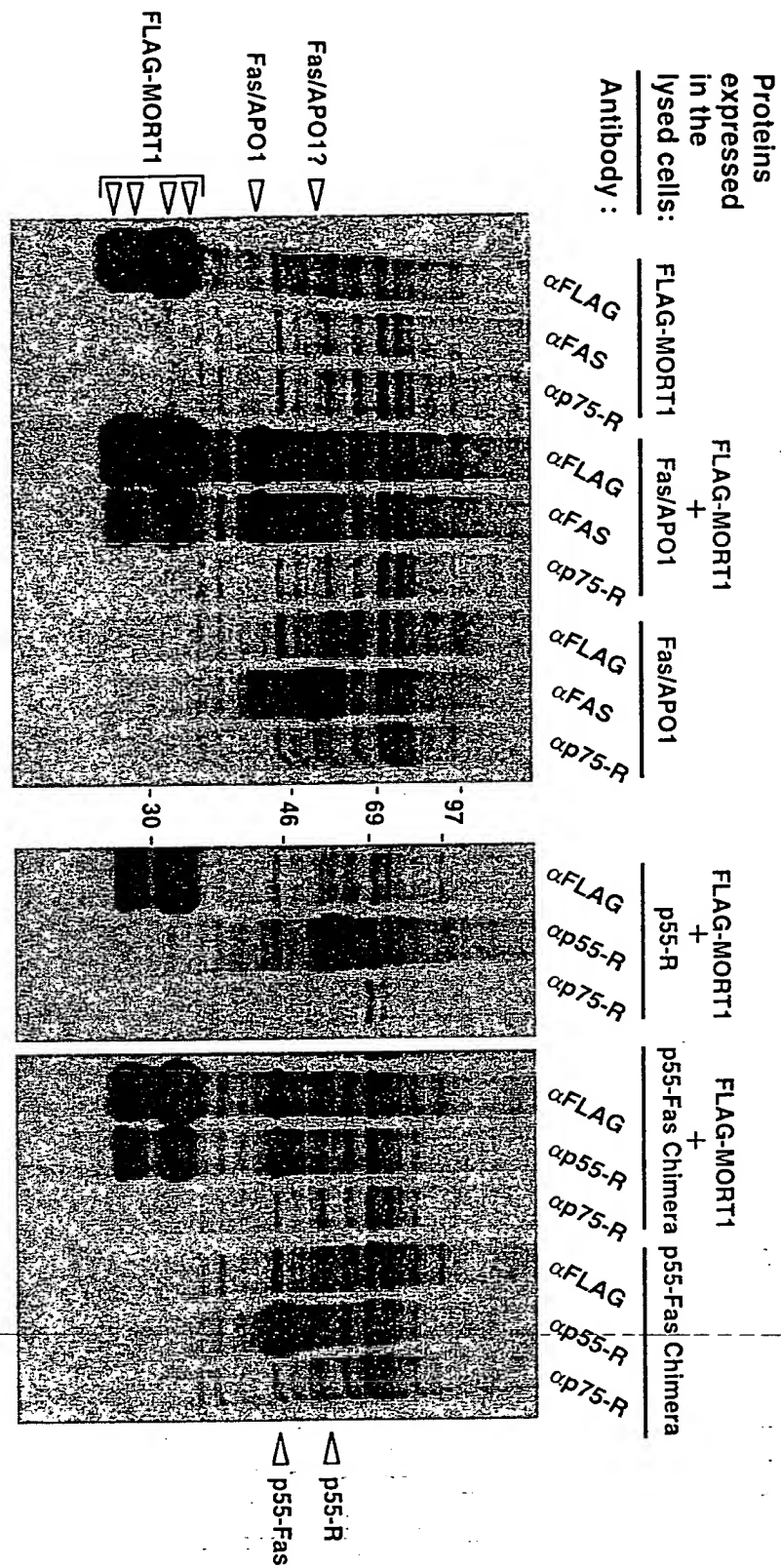


FIGURE 2

28 S →

18 S →

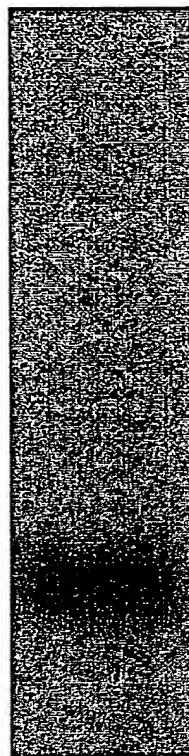


FIGURE 3

1/1
 GTG AAT CAG GCA CCG GAG TGC AGG TTC GGG GGT GGA ATC CTT GGG CCG CTG GGC AAG CGG
 V N Q A P E C R F G G G I L G P L G K R
 61/21
 CGA GAC CTG GCC AGG GCC AGC GAG CCG AGG ACA GAG GGC GCG CCG AGG GCC GGG CCG CAG
 R D L A R A S E P R T E G A R R A G P Q
 121/41
 CCC CGG CCG CTT GCA GAC CCC GCC ATG GAC CCG TTC CTG GTG CTG CTG CAC TCG GTG TCG
 P R P L A D P A M D P F L V L L H S V S
 181/61
 TCC AGC CTG TCG AGC AGC GAG CTG ACC GAG CTC AAG TTC CTA TGC CTC GGG CGC GTG GTC
 S S L S S S E L T E L K F L C L G R V V
 241/81
 AAG CGC AAG CTG GAG CGC GTG CAG AGC GGC CTA GAC CTC TTC TCC ATG CTG CTG GAG CAG
 K R K L E R V Q S G L D L F S M L L E Q
 301/101
 AAC GAC CTG GAG CCC GGG CAC ACC GAG CTC CTG CGC GAG CTG CTC GCC TCC CTG CGG CGC
 N D L E P G H T E L L R E L L A S L R R
 361/121
 CAC GAC CTG CTG CGG CGC GTC GAC GAC TTC GAG GCG GGG GCG GCG GCC GGG GCC GCG CCT
 H D L L R R V D D F E A G A A A G A A P
 421/141
 GGG GAA GAA GAC CTG TGT GCA GCA TTT AAC GTC ATA TGT GAT AAT GTG GGG AAA GAT TGG
 G E E D L C A A F N V I C D N V G K D W
 481/161
 AGA AGG CTG GCT CGT CAG CTC AAA GTC TCA GAC ACC AAG ATC GAC AGC ATC GAG GAC AGA
 R R L A R Q L K V S D T K I D S I E D R
 541/181
 TAC CCC CGC AAC CTG ACA GAG CGT GTG CGG GAG TCA CTG AGA ATC TgG AAG AAC ACA GAG
 Y P R N L T E R V R E S L R I W K N T E
 601/201
 AAG GAG AAC GCA ACA GTG GCC CAC CTG GTG GGG GCT CTC AGG TCC TGC CAG ATG AAC CTG
 K E N A T V A H L V G A L R S C Q M N L
 661/221
 GTG GCT GAC CTG GTA CAA GAG GTT CAG CAG GCC CGT GAC CTC CAG AAC AGG AGT GGG GCC
 V A D L V Q E V Q Q A R D L Q N R S G A
 721/241
 ATG TCC CCG ATG TCA TGG AAC TCA GAC GCA TCT ACC TCC GAA GCG TCC TGA TGG GCC GCT
 M S P M S W N S D A S T S E A S *
 781/261
 GCT TTG CGC TGG TGG ACC ACA GGC ATC TAC ACA GCC TGG ACT TTG GTT CTC TCC AGG AAG
 841/281
 GTA GCC CAG CAC TGT GAA GAC CCA GCA GGA AGC CAG GCT GAG TGA GCC ACA GAC CAC CTG
 901/301
 CTT CTG AAC TCA AGC TGC GTT TAT TAA TGC CTC TCC CGC ACC AGG CCG GGC TTG GGC CCT
 961/321
 GCA CAG ATA TTT CCA TTT CTT CCT CAC TAT GAC ACT GAG CAA GAT CTT GTC TCC ACT AAA
 1021/341
 TGA GCT CCT GCG GGA GTA GTT GGA AAG TTG GAA CCG TGT CCA GCA CAG AAG GAA TCT GTG
 1081/361
 CAG ATG AGC AGT CAC ACT GTT ACT CCA CAG CGG AGG AGA CCA GCT CAG AGG CCC AGG AAT
 1141/381
 CGG AGC GAA GCA GAG AGG TGG AGA ACT GGG ATT TGA ACC CCC GCC ATC CTT CAC CAG AGC
 1201/401
 CCA TGC TCA ACC ACT GTG GCG TTC TGC TGC CCC TGC AGT TGG CAG AAA GGA TGT TTT TGT
 1261/421
 CCC ATT TCC TTG GAG GCC ACC GGG ACA GAC CTG GAC ACT AGG GTC AGG CGG GGT GCT GTG
 1321/441
 GTG GGG AGA GGC ATG GCT GGG GTG GGG GTG GGG AGA CCT GGT TGG CCG TGG TCC AGC TCT
 1381/461
 TGG CCC CTG TGT GAG TTG AGT CTC CTC TCT GAG ACT GCT AAG TAG GGG CAG TGA TGG TTG
 1441/481
 CCA GGA CGA ATT GAG ATA ATA TCT GTG AGG TGC TGA TGA GTG ATT GAC ACA CAG CAC TCT
 1501/501
 CTA AAT CTT CCT TGT GAG GAT TAT GGG TCC TGC AAT TCT ACA GTT TCT TAC TGT TTT GTA
 1561/521
 TCA AAA TCA CTA TCT TTC TGA TAA CAG AAT TGC CAA GGC AGC GGG ATC TCG TAT CTT TAA
 1621/541
 AAA GCA GTC CTC TTA TTC CTA AGG TAA TCC TAT TAA AAC ACA GCT TTA CAA CTT CCA TAT
 1681/561
 TAC AAA AAA AAA AAA AAA AAA

FIGURE 4

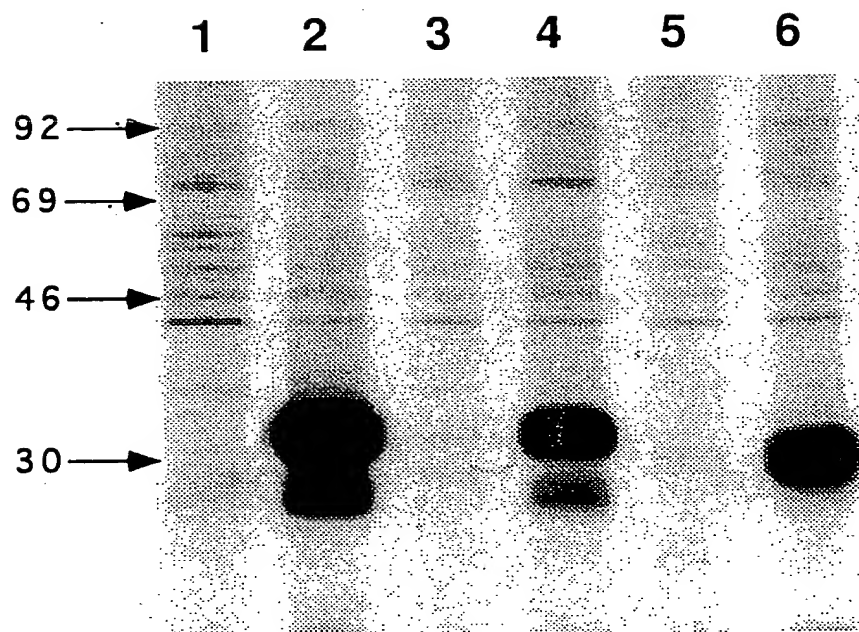


FIGURE 5

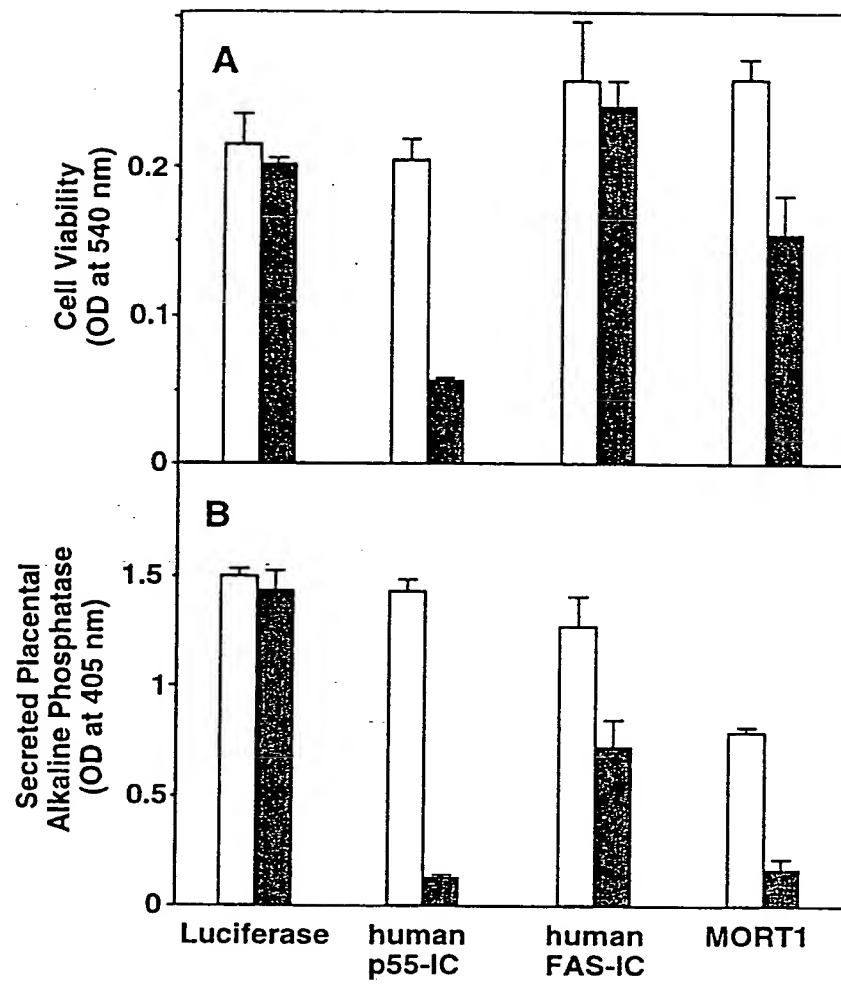


FIGURE 6

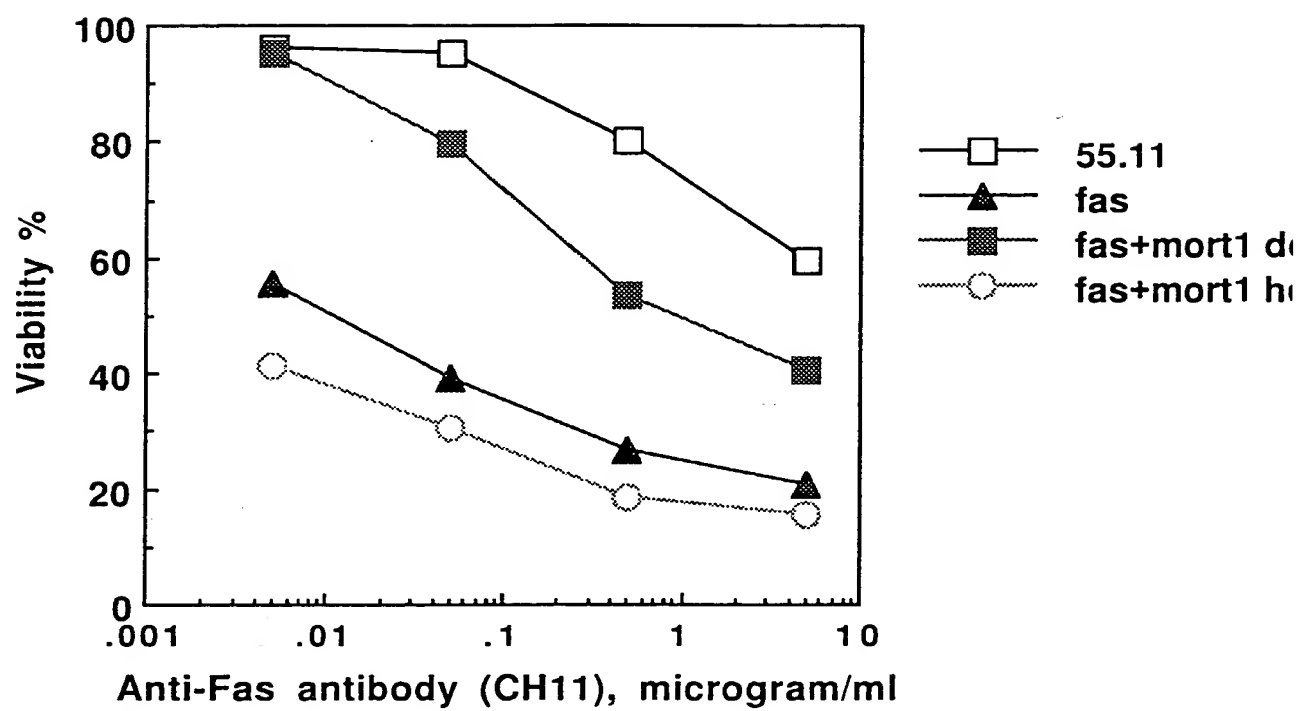


FIGURE 7

ccgccgccgccgccgccacctgcccagacttttctgttccagggtcagcctgtagtgaatcggcc
gctgagcctgaaggaccaacagacgttcgcgcgctctgtgggtctcaaatggcgcaaggtg
gggcgctcactgcagcgaggctgccgggcgctgcgggacccggcgctggactcgctggccta
cgagtacgagcg....

FIGURE 8